

REVIEW

Open Access



Therapeutic potential of autophagy-enhancing agents in Parkinson's disease

Tim E. Moors^{1*} , Jeroen J. M. Hoozemans², Angela Ingrassia¹, Tommaso Beccari³, Lucilla Parnetti⁴, Marie-Christine Chartier-Harlin^{5,6} and Wilma D. J. van de Berg¹

Abstract

Converging evidence from genetic, pathological and experimental studies have increasingly suggested an important role for autophagy impairment in Parkinson's Disease (PD). Genetic studies have identified mutations in genes encoding for components of the autophagy-lysosomal pathway (ALP), including glucosidase beta acid 1 (*GBA1*), that are associated with increased risk for developing PD. Observations in PD brain tissue suggest an aberrant regulation of autophagy associated with the aggregation of α -synuclein (α -syn). As autophagy is one of the main systems involved in the proteolytic degradation of α -syn, pharmacological enhancement of autophagy may be an attractive strategy to combat α -syn aggregation in PD. Here, we review the potential of autophagy enhancement as disease-modifying therapy in PD based on preclinical evidence. In particular, we provide an overview of the molecular regulation of autophagy and targets for pharmacological modulation within the ALP. In experimental models, beneficial effects on multiple pathological processes involved in PD, including α -syn aggregation, cell death, oxidative stress and mitochondrial dysfunction, have been demonstrated using the autophagy enhancers rapamycin and lithium. However, selectivity of these agents is limited, while upstream ALP signaling proteins are involved in many other pathways than autophagy. Broad stimulation of autophagy may therefore cause a wide spectrum of dose-dependent side-effects, suggesting that its clinical applicability is limited. However, recently developed agents selectively targeting core ALP components, including Transcription Factor EB (TFEB), lysosomes, GCase as well as chaperone-mediated autophagy regulators, exert more specific effects on molecular pathogenetic processes causing PD. To conclude, the targeted manipulation of downstream ALP components, rather than broad autophagy stimulation, may be an attractive strategy for the development of novel pharmacological therapies in PD. Further characterization of dysfunctional autophagy in different stages and molecular subtypes of PD in combination with the clinical translation of downstream autophagy regulation offers exciting new avenues for future drug development.

Background

The presence of Lewy bodies (LBs) and Lewy neurites, intraneuronal and axonal inclusions consisting of aggregated α -synuclein (α -syn), is the pathological hallmark of Parkinson's Disease (PD) and Dementia with Lewy Bodies (DLB) [1]. This accumulation of α -syn has been associated with impaired functioning of protein degradation mechanisms [2, 3]. One of the main routes for the intracellular degradation of α -syn is autophagy [4, 5], the

lysosome-mediated catabolic process in which dysfunctional organelles and proteins are degraded in mammalian cells. Autophagy has a pivotal role in the homeostasis of neurons. Protein quality control and organelle recycling are particularly important in these cells, that have abundant oxidative metabolism and generally do not replicate in adult life. Moreover, reduced function of autophagy was demonstrated to cause intracellular accumulation of proteins and neurodegeneration in in vitro and in vivo experiments [6, 7]. The reliance of neurons on proper functioning of the autophagy-lysosomal pathway (ALP) is supported by observations that the brain is often the most severely affected organ in primary lysosomal storage disorders (LSDs) [8].

* Correspondence: t.moors@vumc.nl

¹Department of Anatomy and Neurosciences, Section Clinical Neuroanatomy, Amsterdam Neuroscience, VU University Medical Center Amsterdam, Amsterdam, The Netherlands

Full list of author information is available at the end of the article



After autophagy was implicated in the degradation of α -syn, converging evidence from genetic, pathological and experimental studies has provided indications that the ALP is compromised in PD [9]. In preclinical model systems of PD, the increase of autophagic degradation activity (*autophagic flux*) by rapamycin and lithium, potent autophagy enhancers routinely used in a laboratory setting, resulted in increased clearance of α -syn and neuroprotective effects, which were primarily related to increased autophagy [10]. After these initial findings, similar effects were shown in different preclinical models of PD using other agents that stimulate autophagy, including metformin and trehalose [11]. These studies together suggested a role of autophagy deregulation in the occurrence of different modeled PD-related pathological processes, and suggested that the pharmacological enhancement of autophagy holds promising potential for the development of disease-modifying therapies in PD. However, the selectivity for most of these agents is limited, while upstream ALP components are involved in many other pathways than autophagy. In the context of neurodegenerative disease, in which high-dosage long-term therapies are necessary in order to exert effects in the brain, the broad stimulation of autophagy could therefore result in a wide spectrum of side-effects. Recently, however, several agents that selectively target more downstream ALP components have been developed, which opens up exciting perspectives to combat specific molecular pathogenetic processes relevant for PD.

This study focuses on the emerging role for dysfunctional autophagy in PD, discusses potential molecular targets in the ALP and reviews the evidence obtained in preclinical model systems of PD using autophagy-enhancing strategies, in order to determine the potential of targeted autophagy modulation as disease-modifying therapy in PD.

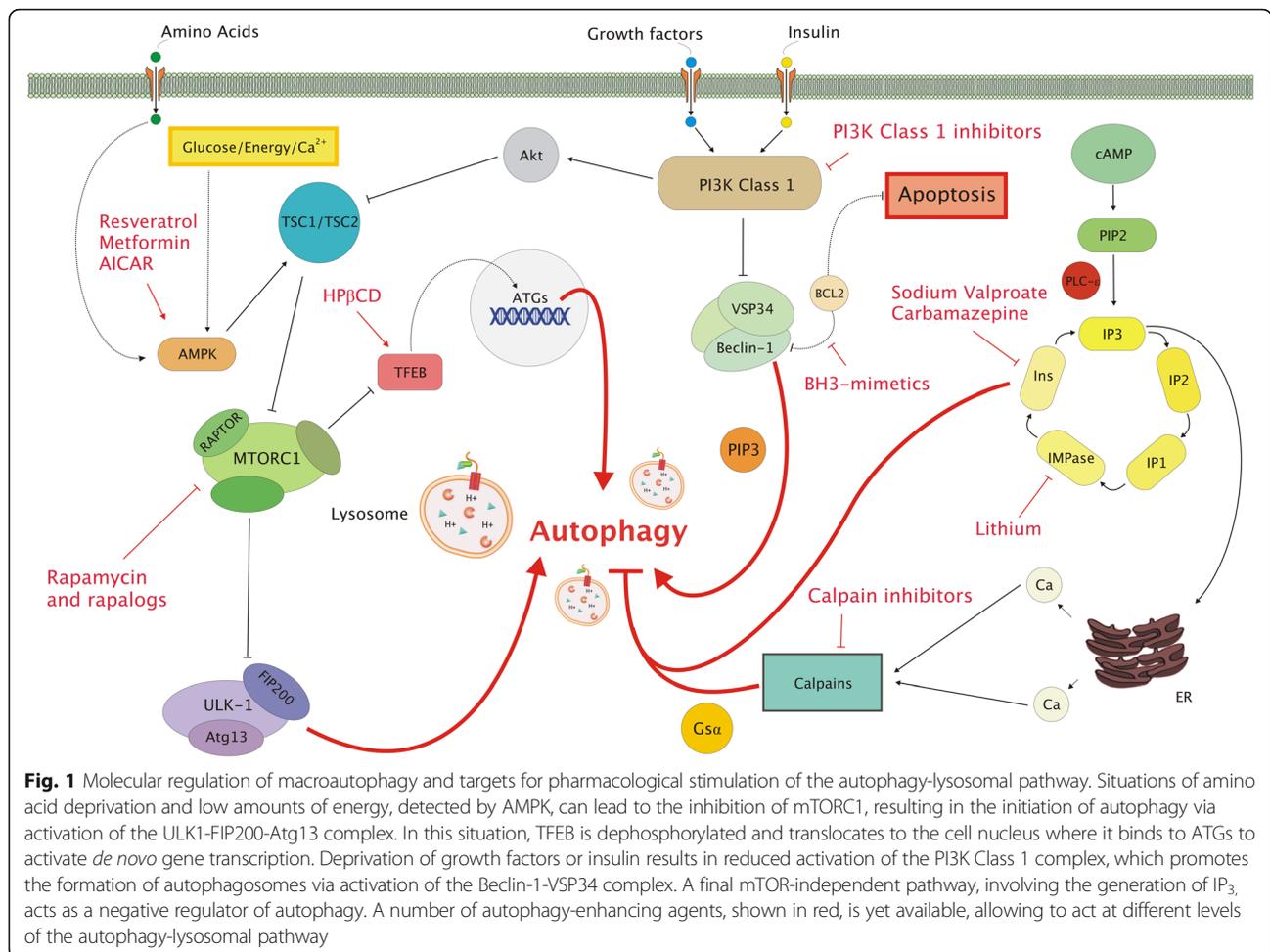
Molecular regulation of autophagy

So far, three types of autophagy have been identified based on routes that differ in the way in which substrates eventually reach the lysosomal lumen: microautophagy, chaperone-mediated autophagy (CMA) and macroautophagy. In microautophagy, cytosolic substrates are engulfed directly by the lysosome, after deformation of the lysosomal membrane, in a non-specific way [12]. Macroautophagy is a degradation pathway that involves the formation, elongation and nucleation of double-membrane organelles - called autophagosomes - by which the substrate is sequestered, before fusion with lysosomes [13]. Finally, chaperone-mediated autophagy (CMA) is a highly specific process in which soluble cytosolic proteins containing a KFERQ-related targeting motif are recognized by a chaperone-complex involving the heat-shock cognate protein of 70 kDa (Hsc70). The substrate is subsequently

translocated to and internalized by the lysosomal-associated membrane protein 2a (LAMP2a) receptor [14]. Under normal conditions, CMA and macroautophagy occur constitutively at low levels, while these processes are triggered under conditions of cellular stress, including starvation, oxidative stress and presence of protein aggregates [13]. While CMA is characterized by its high specificity, macroautophagy has originally been considered a nonspecific bulk degradation pathway. However, a growing number of selective, specialized types of macroautophagy are described which are generally named to the cargo destined for degradation, for instance the targeted degradation of mitochondria (mitophagy), peroxisomes (pexophagy), the endoplasmic reticulum (ER; reticulophagy), ribosomes (ribophagy), lipid droplets (lipophagy) and many more [15]. These specialized forms of macroautophagy have their own degradation cues and specific ubiquitin-dependent or independent autophagy receptors (reviewed in [16, 17]).

Macroautophagy is an evolutionary highly conserved process and its molecular pathways have been well-characterized after the discovery of autophagy-related genes (*ATGs*) in *Saccharomyces cerevisiae* (reviewed in Bento et al. [18]). The mammalian target of rapamycin (mTOR), a 289-kDa serine/threonine kinase, has been identified as a master regulator of macroautophagy, that can be embedded in two protein complexes: mTORC1 or mTORC2. Macroautophagy is negatively regulated by mTORC1, while mTORC2 is primarily involved in regulation of cellular survival and cytoskeletal organization [19]. Situations of amino acid deprivation relieve the direct inhibition of mTORC1 on a protein complex involving UNC-51-like kinase 1 (ULK-1), Atg13 and FIP200, leading to macroautophagy initiation (Fig. 1) [20]. Binding of growth factors or insulin to their corresponding receptors activates the PI3K class 1 protein complex, which can activate mTORC1 via Akt and the tuberous sclerosis complex (TSC1/TSC2 complex) [21]. The activation of PI3K class 1 further results in the inhibition of a macromolecular protein complex including PI3K class 3 (Vps34), Beclin-1 and ATG14L [22], which, when stimulated, promotes autophagosomal membrane nucleation [23]. AMP activated protein kinase (AMPK) detects the intracellular ratio between ATP and AMP and low amounts of energy result in AMPK activation. AMPK activation exerts an inhibiting effect on mTORC1 via the TSC1/TSC2 complex, or in direct phosphorylation of ULK-1, both resulting in the initiation of autophagy [24]. In this way, eukaryotic cells are equipped with a mechanism in which the initiation of autophagy is tightly coupled to cell growth regulation via either inhibition or stimulation of mTOR.

The transcription factor EB (TFEB) has been identified as key regulator of biogenesis and function of lysosomes



and functions downstream of mTORC1 [25, 26]. In resting cells, TFEB is localized at the cytosol, where it interacts with mTORC1 and the lysosomal vacuolar-type ATPase complex. The inhibition of mTORC1 activity results in dephosphorylation of TFEB, which then translocates to the cell nucleus and binds to the lysosome-related genes of the CLEAR network, activating a *de novo* gene transcription [27, 28].

In addition to the mTOR-dependent pathways, macroautophagy can be initiated by a pathway working independent of mTOR (Fig. 1) [29]. This pathway, involving Ca²⁺-calpain-G-stimulatory protein α (G_sα) and cAMP-Epac-PLC-ε-inositol signaling, 1,4,5-triphosphate (IP₃) acts as a negative mTOR-independent regulator of macroautophagy [30]. The generation of IP₃ from PIP₂, which is mediated by PLC-ε, results in the release of calcium from the endoplasmic reticulum (ER). The subsequent activation of calpains leads to cleavage of G_sα and inhibition of macroautophagy. In addition, the conversion of IP₃ induces a chain reaction resulting in the production of Inositol monophosphatase (IMPase) and Inositol (Ins), which also inhibits autophagic processes.

In turn, a decrease of IP₃ leads to reduced calcium release and AMPK activation, which ultimately results in autophagy induction.

Converging evidence for a role of ALP dysfunction in PD

Genetic studies

A substantial amount of recently identified genetic factors has been shown to be involved in or to interact with the ALP, as was reviewed by Gan-Or et al. [31]. Approximately a decade ago, the association between glucosidase beta acid 1 (*GBA1*) mutations and PD was described. Homozygous mutations in the *GBA1* gene, which encodes for the lysosomal hydrolase glucocerebrosidase (GCCase), lead to Gaucher Disease (GD), the most common LSD. Although parkinsonism is a rare feature in patients with GD, several GD patients with parkinsonism had relatives with a typical, late-onset form of PD [32]. After confirmation of this observation in large-scale multicenter studies [33] and meta-analyses [34, 35], the presence of pathogenic heterozygous mutations in this gene is now considered as one of the most important

risk factors to develop PD. It is estimated that the prevalence of PD patients with GBA mutations is 5–10%, while this percentage can be higher in certain populations [36].

Several other lysosomal genes have been associated with PD [37]. Genome-wide association studies have repeatedly reported an association between scavenger receptor class B member 2 (*SCARB2*) with reduced risk to develop PD [31]. *SCARB2* encodes for the lysosomal integral membrane protein type 2 (LIMP2), which interacts with GCCase and is responsible for its transport to the lysosome [38]. The sphingomyelin phosphodiesterase 1 (*SMPD1*) gene encodes for the lysosomal enzyme acid sphingomyelinase (aSMase), which converts sphingomyelin into ceramide. The association of variants of *SMPD1* with increased risk to develop PD has now been repeatedly reported [39–42]. However, *SMPD1* mutations are rare and more studies are needed to determine their significance in PD. Mutations in the *ATP13A2* (*PARK9*) gene, encoding a lysosomal ATPase, cause a rare form of atypical, juvenile-onset autosomal recessive parkinsonism with pyramidal neurodegeneration and dementia - the Kufor-Rakeb syndrome. Finally, some other LSDs have been associated with parkinsonism, including Niemann-Pick type C, Sanfilippo A, GM1 and GM2 gangliosidoses, and Fabry Disease [37]. However, these associations are often based on observations in small patient groups and thus most likely only relevant for very small subpopulations of PD patients.

Apart from mutations in genes that directly encode for lysosomal components, direct and indirect roles have been proposed for numerous other PD-related mutations in the process of autophagy. A rare form of autosomal dominant PD is caused by mutations in the gene encoding vacuolar protein sorting protein-associated protein 35 (*VPS35*), which is involved in endosomal-lysosomal trafficking, a process functionally associated with autophagy [31, 43]. Moreover, numerous autosomal recessive PD genes, including Parkin (*PARK2*), PINK1 (*PARK6*), DJ-1 (*PARK7*) and Fbxo7 (*PARK15*) were found to be implicated in mitophagy, which is the degradation of dysfunctional mitochondria by autophagy [31, 44]. Mutations in the leucine-rich repeat kinase 2 (*LRRK2*) gene are among the most common risk factors for PD together with *GBA1* mutations. *Lrrk2* can both be degraded by CMA [45] and macroautophagy [46], but mutated forms of this protein impair CMA function leading to accumulation of substrates, including α -syn [45]. *Lrrk2* has also been proposed to have a more general role in autophagy as well, as the accumulation of autophagic vacuoles was observed in a cellular model transfected with mutated *LRRK2* [47]. However, the exact role of *Lrrk2* in this process remains elusive [31, 43]. Finally, some proteins encoded by other PD-related genes, including *SNCA* and *MAPT*, were shown to interact

with components of the ALP, thereby modifying the functioning of autophagy [31].

Together, genetic studies have demonstrated that numerous genetic risk factors for PD are directly involved in the functioning of the ALP or mitophagy [31], while for other genetic forms of PD, for instance caused by *LRRK2*, *MAPT* and *SNCA* mutations, functional interactions of their gene-products with components of the ALP were demonstrated. However, it is important to note that the prevalence of most of the mutations described here in PD is low, and that they do not necessarily lead to PD. The genetic studies further suggest that dysfunctional autophagy in PD can have different underlying gene-deficits.

Pathological studies

After accumulation of autophagic vacuoles in the post-mortem SN of PD patients was first described [48], subsequent postmortem studies demonstrated altered expression of numerous ALP components in the PD brain compared to age-matched controls [49]. Increased levels of microtubule-associated protein 1 light chain 3 (LC3), a marker for autophagosomes, have been observed in the SN of PD patients [50] as well as in the temporal cortex of patients with Dementia with Lewy Bodies [51]. Furthermore, lysosomal depletion was indicated by decreased levels of lysosomal-associated membrane protein type 1 (LAMP1) in the SN of PD patients [50, 52]. Lower levels of the CMA markers LAMP2a and the chaperone Hsc70 have further been observed in various regions of the PD brain compared to controls [53, 54], suggesting CMA deregulation. In addition, the deregulation of lysosomal enzymes, in particular GCCase, has been demonstrated in different brain regions [55–57] and CSF [58–60] of PD patients compared to controls, while increased immunoreactivity of the GCCase-interactor LIMP2 was observed in dopaminergic neurons in the SN of PD patients [61]. Altered enzyme activities in the PD brain have not only been observed for GCCase, but also for cathepsin A and cathepsin D [52, 56]. For cathepsin D, increased [56] and decreased [52] activities were reported in the PD cingulate cortex and SN, respectively, while no changes were found in the frontal cortex [62]. Finally, contradictory results were obtained for P-type ATPase (*ATP13A2*) in the frontal cortex of PD and DLB patients, for which both decreased [63] and increased [64] protein levels have been reported compared to controls. In the striatum of PD and DLB patients, increased *ATP13A2* protein levels were found [64].

A deregulation of the ALP in the PD brain was recently demonstrated by transcriptome studies [65–67]. Alterations for numerous autophagy-related processes, including mTOR signaling, PI3K/AKT signaling and 14-3-3 protein signaling, have been reported in postmortem

SN tissue as well as in peripheral blood mononuclear cells (PBMCs) of PD patients [65–67]. The deregulation of the mTOR-dependent pathway in synucleinopathies was underscored by the finding that mTOR protein expression levels were increased in the temporal cortex of patients with DLB in comparison to controls, in particular in neurons displaying α -syn accumulation [68]. Alterations in other upstream autophagy-related proteins were demonstrated in brain tissue of patients with PD and DLB compared to controls [69]. Immunoreactivity of ULK-1, ULK-2, VPS35 and autophagy/Beclin-1 regulator 1 (AMBRA1) was shown within mature LBs [69]. No differences in protein expression levels were observed for these proteins in SN tissue of PD patients, although VPS35 levels were found increased in the temporal cortex of DLB patients [69]. Beclin-1 protein expression levels were found increased in SN tissue [69], but decreased in the cingulate cortex of patients with PD [56]. In the postmortem SN of PD patients, the subcellular localization of TFEB was changed, as TFEB expression in the nuclear department of dopaminergic neurons was significantly decreased in PD patients compared to controls [70]. Moreover, TFEB co-localized with LBs in the same region [70].

In conclusion, pathological studies have shown some evidence for altered ALP protein levels and lysosomal enzyme activity in PD and DLB. However, the reported alterations are generally subtle and sometimes inconsistent, with large overlap between patients and aged controls. Additionally, these pathological studies often also show tremendous variability within the group of PD patients. Together, this may indicate that multiple pathological processes may underpin dysfunctional autophagy in PD or that autophagy is impaired in a subpopulation of PD patients. Moreover, so far, most studies have only focused on the SN of PD patients [49]. As some of the results suggest a differential regulation of autophagy markers between different brain regions, systematic studies that map the distribution of lysosomal and autophagy markers in different stages and molecular subtypes of PD is needed.

Interactions between autophagy and α -syn

Experimental studies have identified the ALP as key player in the clearance of α -syn. Both the ubiquitin-proteasomal system (UPS) and autophagy were found to be implicated in the clearance of wild-type (WT) α -syn in PC12 cell lines, as the inhibition of both processes results in its accumulation [4]. WT α -syn contains the CMA-targeting motif required for Hsc70 recognition, suggesting that its autophagic degradation is associated with CMA activity. Although its clearance can also be partially mediated by macroautophagy, this system may not be of crucial importance in this process [71].

However, a more important role for macroautophagy has been suggested in situations of increased α -syn burden [37], as macroautophagy has been implicated in the clearance of α -syn oligomers [72], as well as mutant [73] and post-translationally modified forms of α -syn, in cellular [74] and yeast [75] models of PD.

Several studies have further shown the possible involvement of lysosomal enzymes in α -syn degradation. First, genetic or pharmacological reduction of GCase activity results in the accumulation of α -syn in in vitro [76, 77], in mice [77], as well as in a transgenic mouse model of GD [78]. Additionally, the depletion of both GCase and cathepsin D lead to enhanced cell-to-cell transmission of α -syn aggregates in vitro, suggesting a role for these enzymes in the spreading and propagation of synucleinopathy [79, 80]. Together, these studies have shown an intimate relation between autophagy, lysosomal enzymes and α -syn turnover. The verification of macroautophagy and CMA contributions in the turnover of different forms of α -syn in vivo, under physiological and pathological conditions, is ongoing [5].

Stimulating macroautophagy in preclinical models of PD

Approximately a decade ago, pioneering studies have explored the chemical stimulation of the ALP in PD model systems using rapamycin and lithium, substances that are routinely used to trigger autophagy in vitro. After this, numerous studies have tested the effects of other autophagy-enhancing agents on different modeled PD-related pathological processes. The following section highlights preclinical studies in which effects of chemical agents in preclinical PD models were primarily associated with increased autophagy (summarized in Table 1).

Pioneering studies: Rapamycin and lithium

The most thoroughly tested macroautophagy-enhancer is rapamycin, a metabolite isolated from the bacterial strain *Streptomyces hygroscopicus*. The drug inhibits the upstream signaling factor mTOR by binding to its intracellular FKBP12 receptor, thereby disrupting the ability of mTOR to form assemblies with RAPTOR and blocking mTORC1 signaling [10, 81]. Rapamycin was able to reduce α -syn accumulation and blocked α -syn induced neurodegeneration in wild-type, A30P, or A53T α -syn expressing PC12 cells [4] and in α -syn-overexpressing mice [68] and rats [70], respectively. Rapamycin further reduced cell death in cell lines treated with 6-OHDA and rotenone [82, 83], and reduced neuronal death in the methyl-4-phenyl-1,2,3,6 tetrahydropyridine (MPTP) mouse model of PD [50, 83]. In addition, rapamycin improved motor function in A53T α -syn overexpressing mice [84] and relieved L-Dopa-induced dyskinesias in 6-OHDA-treated mice and rats without affecting the therapeutic efficacy of

Table 1 Reported effects of autophagy-enhancing agents in preclinical PD models

	Target	Agent	Main effect	PD model	Ref	
mTOR-dependent pathways	AMPK	Metformin	Reduced cell death	Drosophila melanogaster mutated for LRRK2	[95]	
				MPTP mice	[96]	
			Reduced phospho-Ser129 α -syn levels	α -Syn overexpressing SH-SY5Y cells	[98]	
	AMPK	AICAR	Reduced cell death	LRRK2-mutated Drosophila Melanogaster	[95]	
	AMPK (SIRT1)	Resveratrol	Increased α -syn clearance	α -Syn overexpressing PC12 cells	[100]	
			Reduced cell death	Rotenone-exposed SH-SY5Y cells	[100, 101]	
			Improved mitochondrial functioning	Cultured PARK2-mutant fibroblasts	[102]	
	Beclin-1	PREP inhibitor (KYP-2047)	Decreased oligomeric α -syn, increased striatal DA levels	A30P α -syn transgenic mice	[140]	
	Beclin-1	Isorhynchophylline	Increased α -syn clearance	N2a cells transfected for WT, A53T, and A30P α -Syn; Embryonic DA neurons	[141]	
			Increased α -syn clearance/ Reduced α -syn accumulation	WT, A30P, and A53T α -syn expressing PC12 cells	[4]	
				B103 neuronal cells expressing α -syn and Beclin-1	[103]	
				α -Syn-transgenic mice	[68]	
				α -Syn-transgenic rats	[70]	
	mTORC1	Rapamycin and Rp analogues (CCI-779, RAD001 and AP23573)	Reduced phospho-Ser129 α -syn levels	α -Syn overexpressing SH-SY5Y cells	[98]	
			Reduced cell death	Rotenone-exposed SH-SY5Y cells	[82, 92, 93]	
				6-OHDA and MPP+ treated PC12 cells	[83]	
					MPTP mice	[50, 83]
					α -Syn-transgenic mice	[68]
					α -Syn-transgenic rats	[70]
					Drosophila melanogaster mutated for PINK-1 and Parkin	[87]
				Improved motor function, reduced synaptic injury	A53T- α -Syn transgenic mice	[84]
				Reduced levodopa-induced dyskinesia	6-OHDA mice; 6-OHDA rats	[85, 86]
				Reduced mitochondrial dysfunction	Rotenone-exposed SH-SY5Y cells	[82, 92]
			Drosophila melanogaster mutated for PINK-1 and Parkin	[87]		
TFEB	2-HP β CD	Increased α -syn clearance	H4 human neuroglioma cells transfected for α -syn	[143]		

Table 1 Reported effects of autophagy-enhancing agents in preclinical PD models (*Continued*)

mTor-independent pathways	IMPase	Lithium	Increased clearance of A53T and A30P α -syn	PC12 cells expressing A53T and A30P α -syn	[88]
			Reduced apoptosis and mitochondrial dysfunction	Rotenone-exposed SH-SY5Y cells	[92, 93]
			Improved motor function, increased viability DA cells in the SN, decreased loss of DOPAC	MPTP mice (combined treatment with lithium and sodium valproate)	[94]
	Ins	Sodium Valproate	Reduced apoptosis and mitochondrial dysfunction	Rotenone-exposed SH-SY5Y cells	[92]
			Improved motor function, increased viability DA cells in the SN, decreased loss of DOPAC	MPTP mice (combined treatment with lithium and sodium valproate)	[94]
	Ins	Carbamazepine	Reduced apoptosis and mitochondrial dysfunction	Rotenone-exposed SH-SY5Y cells	[92]
	SLC2A transporters	Trehalose	Reduced cell loss	Rotenone-treated rats and PC12 cells	[107]
				MPTP mice; A53T α -Syn overexpressing rats	[108, 109]
			Increased α -syn clearance	PC12 cells overexpressing WT and A53T α -Syn	[104, 106]
				NB69 human neuroblastoma cells	[105]
			Rotenone-treated PC12 cells	[107]	
			A53T α -Syn overexpressing rats	[109]	
			A53T α -Syn overexpressing mice	[110]	
			Reduced motor deficits	MPTP mice; A53T α -Syn overexpressing rats	[108, 109]
			Reduced neuroinflammation	MPTP mice	[108]
Unknown			SMERs (SMER 10, 18 & 28)	Increased A53T α -syn clearance	PC12 cells expressing A53T α -syn
Unknown	Latrepidine	Increased α -syn clearance	Saccharomyces cerevisiae, SH-SY5Y cells expressing α -syn and WT mice	[115]	
		Decreased cell death	Saccharomyces cerevisiae expressing α -syn	[115]	
Unknown	Spermidine	Reduced motor dysfunction, increased lifespan; Reduced neuronal cell loss	Drosophila melanogaster expressing α -syn; Caenorhabditis elegans expressing α -syn	[118]	
Unknown (polyphenols)	Curcumin	Reduced α -syn accumulation	SH-SY5Y Cells expressing WT and A53T α -syn	[119]	
	Kaempferol	Reduced ROS, apoptosis, and mitochondrial dysfunction	Rotenone-exposed SH-SY5Y cells, mouse primary neuronal culture	[120]	
	<i>C. album</i> polyphenol fractions	Reduced α -syn accumulation; reduced ROS	Hu neuroglioma H4 cells expressing α Syn; Yeast cells expressing α Syn	[121]	
Unknown	Nilotinib	Increased α -syn clearance; improved motor function	Mice expressing A53T α -synuclein; mouse primary cortical neurons	[122, 123]	
		Reduced cell death	Mice expressing A53T α -synuclein	[122]	

Table 1 Reported effects of autophagy-enhancing agents in preclinical PD models (*Continued*)

Lysosomes	GCase	Ambroxol	Restoration of lysosomal function; increased GCase activity	GBA1 mutant fibroblasts	[150, 151]
			Reduction of oxidative stress	GBA1 mutant fibroblasts	[150]
	GCase	Isofagomine	Improved motor performance, increased α -syn clearance, reduced neuroinflammation	WT- α -syn overexpressing mice	[159]
	Lysosome	Acidic Nanoparticles	Restoration of lysosomal function; reduced DA cell loss	Cultured ATP13A2 and GBA-mutant fibroblasts; MPTP mice	[145]

Abbreviations: Ref reference number, 2-HP β CD 2-Hydroxypropyl- β -cyclodextrin, α -syn α -synuclein, DA dopaminergic, DOPAC 3,4-Dihydroxyphenylacetic acid, ROS reactive oxidative stress, WT wild-type, 6-OHDA 6-hydroxydopamine, MPTP 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine, UPS ubiquitin-proteasomal system

L-Dopa [85, 86]. Finally, rapamycin showed neuroprotective effects in *Drosophila melanogaster* mutated for PINK-1 and Parkin [87]. However, rapamycin interferes in numerous other pathways than autophagy, while prolonged treatment with rapamycin can lead to the inhibition of mTORC2, thereby possibly stimulating other important cellular pathways, for instance influencing cell survival mechanisms. In addition, the use of rapamycin has been associated with a wide spectrum of side-effects, including oral and respiratory infections, stomatitis, leukopenia, hypertriglyceridemia, hypercholesterolemia and immunosuppression [10].

Lithium, which is in use as a mood stabilizer in the treatment of bipolar disorders, is able to induce autophagy in an mTOR-independent manner by direct non-competitive inhibition of IMPase [88]. The neuroprotective effects of lithium in in vivo and in vitro models have repeatedly been reported [89]. However, lithium interferes with many other cellular pathways as well, and numerous mechanisms of action have been proposed for this drug [90]. The use of lithium in bipolar disorders revealed that the drug is associated with a wide spectrum of dose-dependent side-effects, for instance including fine hand tremor, hypothyroidism, hypercholesterolemia, hyperparathyroidism, and hypercalcemia [91]. In preclinical PD models, lithium prevented accumulation of α -syn in PC12 cells expressing A53T and A30P α -syn [88] and protected against rotenone-induced neurotoxicity and cell death via the induction of autophagy in different cell lines [92, 93]. Two other mood-stabilizing agents - sodium valproate and carbamazepine—were demonstrated stimulate autophagy independently from mTOR via Ins, which is downstream of IMPase [92]. Both sodium valproate and carbamazepine were shown to decrease rotenone toxicity and to induce autophagy in SH-SY5Y cells [92]. Moreover, combined treatment of sodium valproate with lithium alleviated motor impairments in MPTP mice, while it protected SN dopaminergic neurons [94]. However, similar to lithium, these agents are non-selective for autophagy and acts on many other cellular pathways as well, possibly resulting in a numerous unwanted effects.

Together, preclinical studies using rapamycin and lithium showed beneficial effects on different modeled PD-related pathological processes, including α -syn aggregation, mitochondrial dysfunction and oxidative stress, which were associated with increased autophagy. Both agents affect many other cellular processes than autophagy, and their side-effect profile makes them unsuited for prolonged high-dosage therapies. However, these pioneering studies have provided important insights into the role of autophagy in PD-related pathological processes, and demonstrated the potential of autophagy-enhancing strategies in experimental settings.

Other targets for macroautophagy enhancement

AMPK

Activation of AMPK, the signaling factor upstream of mTORC1, leads to the inhibition of mTORC1 and initiation of autophagy. Numerous agents are associated with the activation of AMPK, including metformin, 5-aminoimidazole-4-carboxamide ribonucleotide (AICAR) and resveratrol. The administration of metformin, which is used in Diabetes Mellitus, showed neuroprotective effects in in vitro and in two in vivo models of PD: *drosophila* with mitochondrial dysfunction and MPTP mice [95–97]. Further, metformin decreased Ser-129 phosphorylated α -syn levels in α -syn overexpressing SH-SY5Y cells [98]. Similar results have been reported using AICAR, an drug used in the treatment of acute lymphoblastic leukemia [95–97]. Resveratrol activates AMPK after interaction with its direct target SIRT1 and protective effects of this compound have been reported in different in vitro and in vivo models of PD [99–102], some of which were associated with increased autophagy [100–102].

Trehalose is a disaccharide that inhibits members of the SLC2A (also known as GLUT) family of glucose transporters, leading to an AMPK-dependent - and mTOR-independent - increase in autophagy [103, 104]. The beneficial effects of this drug for cell survival and α -syn clearance have been shown and associated to increased autophagy in different cell lines [104–106] as

well as in multiple *in vivo* models [107–110]. As trehalose is digested by trehalase into glucose in the small intestine, liver and kidney, its bioavailability in the brain may be limited. Recently, two novel trehalase-indigestible and autophagy-inducing disaccharides were developed, which were shown to reduce polyglutamine aggregation *in vitro*, highlighting the anti-aggregation properties of these agents [111], which, to our knowledge, have not been tested in preclinical models of PD, yet. In sum, AMPK-dependent increase of autophagy showed beneficial effects on different modeled pathological processes in PD. However, importantly, as cellular energy sensor, AMPK is involved in various other key cellular processes, including cell proliferation and survival. Therefore, the global manipulation of its function is likely to induce unwanted effects, and limits its therapeutic potential in the context of PD.

Other macroautophagy-enhancing agents

A chemical screening approach has resulted in the discovery of three small-molecule enhancers of rapamycin (SMERs): SMER10, SMER18 and SMER28 [112]. These agents induced autophagy and enhanced clearance of mutant A53T α -syn in a PC12 cell line in an mTOR-independent way [112]. Although SMERs have shown promising potential in PD model systems, their specific biochemical pathways remain to be characterized [113]. The antihistamine latrepirdine was shown to enhance autophagy [114] and to attenuate accumulation of α -syn both *in vivo* and *in vitro* systems [115]. Latrepirdine entered phase 3 clinical trials for both Alzheimer's disease (AD) and HD [116], in which was well-tolerated without significant toxicity it failed to show efficacy compared to placebo (more information available on: <https://clinicaltrials.gov/>). Although latrepirdine was suggested to act preferentially on α -syn [117], it probably intervenes in many cellular processes, while its biochemical pathways are currently unclear [116]. Finally, numerous studies have associated beneficial effects in preclinical models of PD of other compounds, for instance including spermidine [118], different dietary polyphenols other than resveratrol [119–121] and tyrosine kinase inhibitors [122, 123], with increased macroautophagy. However, the exact mechanisms of action for these compounds require further investigation, and these agents may affect many other cellular processes other than autophagy.

Macroautophagy: a double-edged sword

Together, studies using macroautophagy-enhancing agents in preclinical models of PD have demonstrated that the broad stimulation of autophagy can result in the alleviation of different modeled PD-related pathological processes, such as cell death, α -syn aggregation, oxidative stress and mitochondrial dysfunction (Table 1).

However, the clinical translation of these findings is problematic, given that the selectivity of these agents for autophagy is limited, while upstream ALP signaling proteins are involved in many pathways other than autophagy - for instance including apoptosis, cell growth, and immune responses. This lack of selectivity may limit the therapeutic potential of broad autophagy stimulation, as it may result in unwanted effects. Long-term effects and side-effects of broad stimulation of macroautophagy are currently unknown, and more insight in these effects is of crucial importance before application of prolonged high-dose therapies in a clinical setting. In particular, the intimate relation between macroautophagy and apoptosis should be carefully considered. Although macroautophagy and apoptosis comprise two distinct cellular processes, various shared components and mechanisms have been identified by which autophagy and apoptosis regulate each other [124].

Another important danger of broad macroautophagy enhancement is the complex dual role for autophagy in specific contexts. For example, autophagy can have tumor-suppressive, neutral, or tumor-promoting effects in different types and stages of cancer [125]. Although autophagy can act as tumor suppressor, for instance by the removal of damaged organelles, autophagy stimulation can also promote the growth of established cancers in certain contexts [125, 126]. Indeed, excessive stimulation of macroautophagy under specific circumstances has been associated with detrimental effects. First, the observation of a specific type of cell death characterized by extensive autophagic vacuolization of the cytoplasm in PD postmortem brain tissue [48] has led to the highly debated concept of 'autophagic cell death'. In various *in vitro* model systems of PD, autophagic cell death was reported in cells treated with excessive levels of dopamine [127], A53T α -syn [128] and oxidative stress [129]. Moreover, treatment with rapamycin in presence of oxidative stress reduced cell viability [129], while the activation of AMPK has been reported to induce cell death in different *in vitro* models of PD [130].

The activation of macroautophagy has further been associated with the shortening of neurites in a 6-OHDA mouse model, while the indirect activation of mTOR by the inhibition of Akt was able to suppress the retrograde axonal degeneration [131]. Autophagy was further associated with neurite degeneration in LRRK2-mutant SH-SY5Y cells [47] as well as in mouse superior cervical ganglion neurons [132]. Overexpression of both WT and A53T α -syn in SH-SY5Y and PC12 cell lines resulted in a gradual increase in toxicity, mediated by impairment of CMA [133]. The induction of macroautophagy in these models led to neuronal death, indicating that a compensatory upregulation of macroautophagy under the circumstances of increased α -syn burden can have

detrimental effects [133]. Furthermore, the activation of macroautophagy in primary cortical neurons overexpressing A53T α -syn caused mitochondrial destruction and loss, as well as neuronal degeneration [134]. Finally, another form of autophagy-dependent cell death – autosis – was recently described, although its exact mechanisms remain unclear [135].

Taken together, these experimental findings demonstrated that activation or stimulation of macroautophagy can have detrimental effects under specific circumstances. Maintaining the balance between protective and detrimental effects is therefore of vital importance in therapeutic approaches to stimulate broad autophagy [136], which implies extensive knowledge about dosage and timing for such therapies and may limit their current therapeutic applicability.

Selective targeting of ALP components

Beclin-1

The activation of Beclin-1 leads to autophagosome formation and initiation of autophagy, independent from mTOR (Fig. 1). Gene-transfer induced overexpression of Beclin-1 was able to reduce accumulation of α -syn in α -syn overexpressing mice [137] and PC12 cells [138], suggesting that the Beclin-1 pathway may be a viable therapeutic target in PD. Although numerous drugs have been associated with increased Beclin-1 activity, including tamoxifen, Beclin-1 expression mimetics (BH3 mimetics) and the peptide Tat-Beclin-1, only few studies tested pharmacological Beclin-1 stimulation in PD models [139–141]. One study showed protective effects of tamoxifen, a drug widely used in the treatment of breast cancer, in a MPP⁺ rat model, in which it suppressed radical generation [139]. However, this effect was not attributed to increased autophagy. Another study reported that the inhibition of prolyl oligopeptidase (PREP) resulted in a Beclin-1-dependent increase of autophagy together with a reduction of α -syn aggregates in cell models and α -syn transgenic mice [140]. Finally, isorhynchophylline, a natural alkaloid, was reported to promote the clearance of WT, A53T and A30P α -syn monomers, as well as oligomeric and aggregated forms of α -syn in neuronal cell lines, in which its autophagy-enhancing activity was shown to be Beclin-1-dependent [141]. As the BH-3 domain of Beclin-1 interacts with the pro-survival BCL-2 family members, it was expected that binding of Beclin-1 to BCL2 would induce apoptosis, potentially resulting in undesired side-effects [126]. However, overexpression of Beclin-1 does not result in induction or enhancement of apoptotic processes, which suggested that the functional interaction of Beclin-1 and BCL-2 is unidirectional, resulting in inhibitory effects on autophagy without modifying apoptotic processes [126].

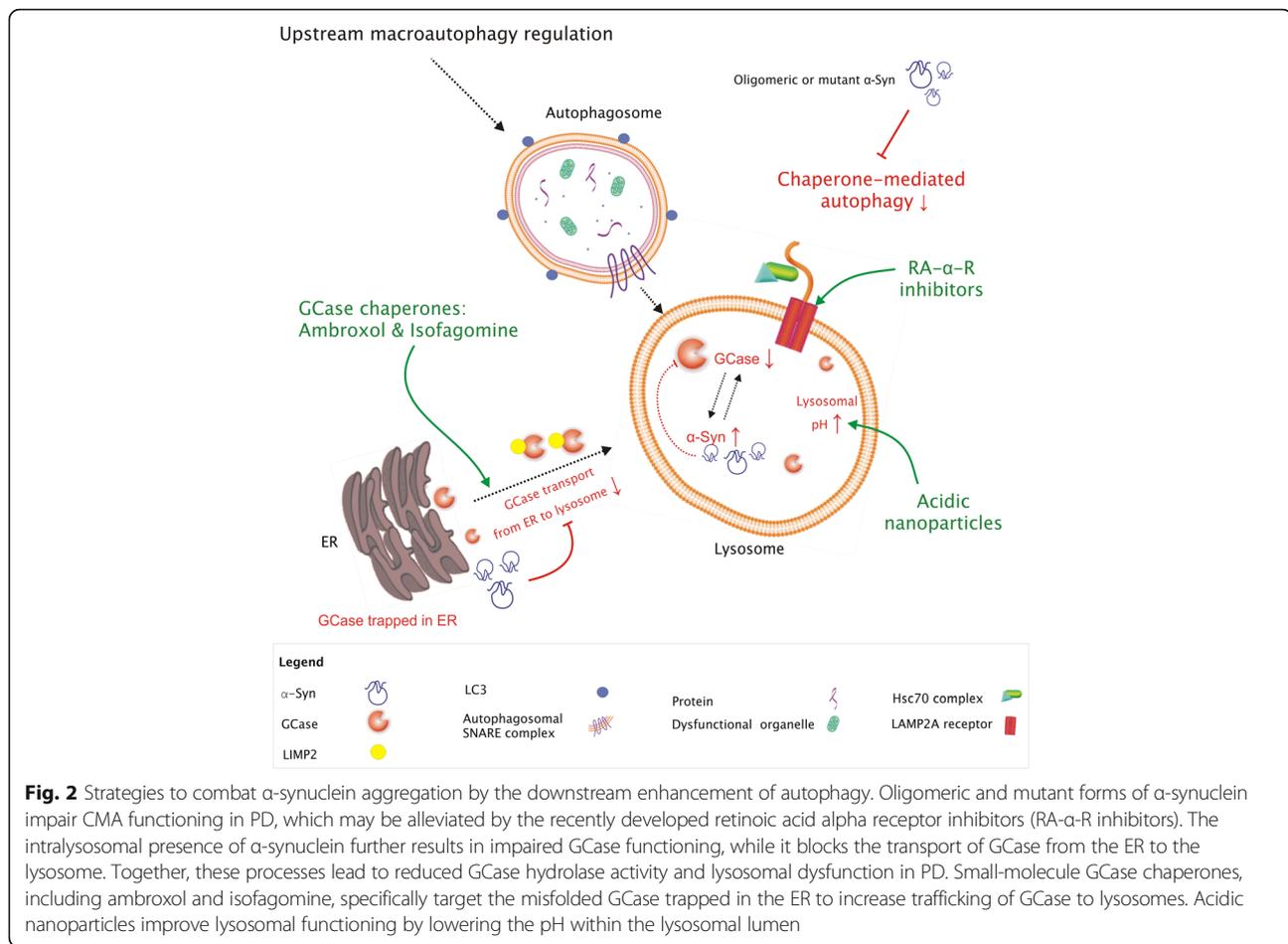
Macroautophagy stimulation downstream of mTOR

As the main transcriptional regulator of the CLEAR network, which coordinates biogenesis and function of lysosomes, TFEB is a crucial link between the upstream signaling pathways that regulate macroautophagy and lysosomes [26]. Therefore, TFEB forms an attractive target to stimulate macroautophagy downstream of mTOR, by intervening at the transcriptional level (Fig. 2). The first approaches to stimulate TFEB in PD models have yielded promising results. Overexpression of TFEB was demonstrated to eliminate α -syn oligomers and to rescue midbrain dopamine neurons from α -syn toxicity in α -syn overexpressing rats [70]. In addition, delayed activation of TFEB via inhibition of mTORC1, achieved using a derivative of rapamycin (CCI-779), alleviated α -syn pathology and was associated with increased autophagic markers and the nuclear translocation of TFEB [70]. 2-Hydroxypropyl- β -cyclodextrin enables the chemical activation of TFEB [142], and was recently shown to promote autophagic clearance of α -syn [143]. These findings provided a mechanistic link between TFEB function and α -syn aggregation, highlighting the role of TFEB in synucleinopathy and the potential of TFEB-activating strategies to combat α -syn aggregation in PD.

Direct targeting of lysosomes

The modulation of lysosomes has recently emerged as an attractive strategy to selectively stimulate the ALP in PD. Recent work showed that acidic nanoparticles were able to stimulate lysosomal degradation by the lowering of lysosomal pH [144], reverting lysosomal dysfunction in three different PD genetic models, including ATP13A2-mutant cells and GBA-mutant cells [145]. Moreover, acidic nanoparticles attenuated dopaminergic cell death in MPTP-treated mice, providing evidence for the feasibility of such strategies *in vivo* [145]. These findings highlight the potential of pharmacological interventions on the lysosomal system.

Another interesting approach is the direct targeting of specific lysosomal enzymes such as GCase to stimulate the intralysosomal degradation of α -syn. Restoration of GCase expression by an adeno-associated virus reduced α -syn aggregation and, interestingly, its expression in the hippocampus was able to reverse cognitive impairment in a GD mouse model [78, 146, 147]. As results from the enzyme-replacement therapies in GD show that GCase does not cross the blood-brain barrier (BBB), the development of small-molecule chaperones to correct the folding of GCase – thereby enhancing GCase activity and lysosomal function – is currently a topic of great interest [148]. Small-molecule chaperones specifically target the misfolded GCase trapped in the ER, stabilizing the active form of the enzyme and increases trafficking of GCase to lysosomes (Fig. 2) [149].



The pharmacological chaperone ambroxol, which has been in use for airway diseases, was shown to increase GCCase activity and reduce oxidative stress in PD fibroblasts carrying GBA1 mutations [150]. Furthermore, ambroxol was shown to restore Cathepsin D, LIMP2 and Saposin C levels [151], which are essential for the proper functioning of GCCase [149, 152]. In this study, ambroxol acted preferentially on lysosomal dynamics, as it did not interfere with Parkin levels or proteasome 20S activity [151]. Although ambroxol was initially expected not to have CNS effects at clinically used doses [153], the drug was reported to cross the BBB in mice [154]. Recently, a pilot study carried out in neuronopathic GD patients showed that high-dose oral ambroxol could cross the BBB, being also well-tolerated, in addition to enzyme-replacement therapy. In these patients, ambroxol increased lymphocyte GCCase activity, reduced CSF glucosylsphingosine levels and was associated with improvements in myoclonus, seizures and pupillary light reflex dysfunction. Accordingly, ambroxol represents an interesting candidate waiting for clinical trials in PD. Other small-molecule chaperones such as AT2101 (isogomine) as well as histone deacetylase inhibitors

have been reported to increase GCCase levels in GD mouse models and fibroblasts [155–158], and are currently tested in the context of PD. Additionally, the administration of isogomine to human WT- α -syn overexpressing mice improved motor performance, reduced α -syn immunoreactivity in nigral dopaminergic neurons and reduced inflammatory microglial activation [159].

Together, small chaperone activity-enhancers of GCCase were demonstrated to improve lysosomal function and enhance α -syn clearance in preclinical models of PD. However, a disadvantage of these chaperones is that they inhibit GCCase activity by binding the catalytic site of the enzyme, suggesting that the balance between chaperone functions and inhibitory activity should be carefully considered when using these compounds [149]. Therefore, molecular chaperones that do not bind the catalytic site of GCCase are required to exert maximal effects. Recently, a high throughput screening has led to the identification of pyrazolopyrimidine derivatives that did not inhibit GCCase, but still facilitated its translocation to the lysosome [160]. These agents were able to enhance GCCase activity, reduce substrate accumulation and normalize oxygen species production in a macrophage model of

GD [161]. Another recent study identified a small-molecule noninhibitory chaperone of GCase (NCGC607) by high-throughput screening [162]. This chaperone was demonstrated to restore GCase activity and protein levels and reduced substrate levels, while it reduced α -syn levels in iPSC-derived dopaminergic neurons [162]. This study underscores the potential of small-molecules targeting GCase activity and protein to prevent or reduce accumulation of α -syn levels in vitro.

Targeting chaperone-mediated autophagy

CMA impairment in PD has been demonstrated by the lower levels of the CMA markers LAMP2a and the chaperone Hsc70 in various regions of the PD brain compared to controls [53, 54], while a central role for this system is proposed in the degradation of WT α -syn [5]. An attractive alternative to the stimulation of macroautophagy may therefore be the downstream targeting of CMA components, at the level of LAMP2a and Hsc70. The overexpression of LAMP2a has been induced in human SH-SY5Y cells, rat primary cortical neurons in vitro and nigral dopaminergic neurons in vivo [163]. In these different models, overexpression of LAMP2a decreased α -syn accumulation and protected against the α -syn-induced dopaminergic degeneration [163]. Interestingly, retinoic acid alpha receptors (RA- α -Rs) were recently identified as CMA inhibitors and synthetic derivatives of all-trans-retinoic acid were developed in order to neutralize this effect [164]. These derivatives were able to specifically stimulate CMA without a compensatory blockage of macroautophagy, while LAMP2a was identified as one of their downstream targets [164]. However, so far, no studies have reported the chemical modulation of CMA in preclinical models of PD yet.

Modulation of mitophagy

The involvement of different PD-related genes in mitophagy suggest a role for impaired mitochondrial clearance in PD. By stimulating the removal of damaged mitochondria, the therapeutic stimulation of mitophagy may therefore exert neuroprotective effects. However, pharmaceutical agents that selectively increase mitophagy are currently lacking. The compounds used in vitro to trigger mitophagy, such as trifluorocarbonyl cyanide phenylhydrazone (FCCP) and antimycin/oligomycin combinations, are toxic and therefore not considered for therapy [165]. The p62-mediated mitophagy inducer (PMI) may be a promising chemical candidate, as it is suggested to enhance endogenous levels of mitophagy [166]. However, no effects of this agent in preclinical models of PD have been described yet. More insights into the complex dynamics of mitochondria and the identification of targets able to specifically stimulate mitochondrial clearance may allow the development

of innovative neuroprotective therapies for PD and other neurodegenerative disorders.

Autophagy-related micro-RNAs

In recent years, an important role has been suggested for micro-RNAs (miRs) in the post-translational regulation of autophagy at various stages of the ALP [167], which interconnect autophagy with other cellular signaling systems. A recent study showed the down-regulation of miR-124, which is predicted to regulate 52 genes of the ALP in a MPTP mouse model [168, 169], while the delivery of an miR-124 agomir attenuated lysosomal depletion and cell death [169]. A different study demonstrated an important role for different miRs in the regulation of CMA markers LAMP2a and Hsc70 [170]. Interestingly, transfection of these miRs resulted in decreased LAMP2a and Hsc70 protein levels as well as in significant α -syn aggregation [170]. Moreover, CMA-related miRNAs were significantly increased in SN and amygdala of PD patients [170]. A better understanding of the interaction between miRs and ALP components and their role in PD may contribute to valuable insights in autophagy regulation and the cross-talk of autophagy with other cellular processes, and possibly to future autophagy-enhancing strategies [169, 170].

The therapeutic value of autophagy enhancers in PD: considerations and perspectives

The identification of autophagy as key player in the degradation of α -syn has suggested the potential of autophagy-enhancing strategies as disease-modifying therapy to combat α -syn aggregation in PD. Indeed, stimulation of autophagy by known autophagy enhancers such as rapamycin and lithium, resulted in increased α -syn clearance in different α -syn overexpressing preclinical model systems of PD. Moreover, interestingly, increased autophagy levels were associated with neuroprotective effects in toxic model systems of PD, as pharmacological autophagy enhancement ameliorated cell death and mitochondrial dysfunction, possibly mediated by the increased clearance of damaged mitochondria (Table 1). Together, pioneering studies have highlighted potential effects of autophagy-enhancement on different modeled PD-related pathological processes. However, selectivity of the early autophagy-enhancing agents and their targets is limited and broad macroautophagy stimulation may result in a wide spectrum of dose-dependent side-effects, making it less suited for high-dosage long-term therapies. In addition, the broad and excessive stimulation of macroautophagy under specific, yet not fully characterized, circumstances can have detrimental effects, including autophagic cell death.

The recent development of agents selectively targeting ALP core components - including TFEB, lysosomes and GCase—as well as CMA factors such as

LAMP2a, enables more specific targeting of pathogenetic processes, and may harbor more perspective for the development of disease-modifying therapies in PD than the broad activation of macroautophagy. CMA is a highly specific process that is increasingly recognized as a key system in the degradation of WT α -syn [37], which highlights the demand for the discovery and development of chemical substances able to specifically stimulate CMA traffic. Targeting lysosomes or GCCase by acidic nanoparticles and small-molecule chaperones, respectively, has emerged as attractive strategy to encounter lysosomal depletion in PD, representing an area of intensive investigation. Moreover, the modulation of downstream ALP components and CMA could lead to more controlled activation of macroautophagy, as substantial cross-talk takes place between CMA and macroautophagy [37, 171]. In the study by McNeill et al., for instance, the GCCase chaperone ambroxol exerted marked effects on TFEB expression, which suggested that the protective effects were partially mediated by increased macroautophagy [150]. The identification of the exact mechanisms underlying the cross-talk between autophagy pathways, lysosomes and GCCase may allow to identify potential dangers and to determine optimal efficacy of downstream autophagy-enhancing approaches.

Establishing the patient selection, optimal dosage, and timing for maximal therapeutic efficiency will be crucial next steps in the design of autophagy-enhancing therapies. Timing of the autophagy-enhancing therapies may be of particular importance, given the hypothesized differential roles for proteolytic systems in α -syn degradation in different stages of aggregation [37]. In vitro experiments suggested that a compensatory upregulation of macroautophagy under the circumstances of increased α -syn burden can have detrimental effects [133]. LB-like aggregates were further shown to impair macroautophagy by blocking clearance of autophagosomes [172], indicating that excessive macroautophagy stimulation in situations of advanced synucleinopathy may result in autophagic cell death. A better understanding of the role of ALP dysfunction in different stages of PD is therefore essential for the design of future disease-modifying therapies.

Pathological and genetic studies further suggested that various molecular and genetic deficits can underlie dysfunctional autophagy in PD. Therefore, the nonspecific stimulation of autophagy may have differential outcomes depending on the molecular subtype of PD. For instance, upregulation of autophagy in the context of lysosomal dysfunction, in which lysosomal clearance of autophagosomes is impaired, may lead to further accumulation of autophagosomes. More insight into the contribution of dysfunctional autophagy to the progression of these different genetic and molecular subtypes of PD is of key

importance in order to identify potential dangers and to develop autophagy-enhancing therapies that target subtype-specific key pathological processes. The development of relevant model systems for these subtypes of PD is therefore urgently required. Finally, to monitor therapeutic response of autophagy-enhancing agents, the development of novel assays that reliably measure dynamics in autophagic flux in vivo are highly desired [11].

Conclusion

In conclusion, pioneering studies using macroautophagy-enhancing agents in preclinical models have provided important knowledge about the role of autophagy in PD and highlighted the promising perspective of autophagy enhancement in this disease. The neuroprotective actions of broad autophagy enhancement were demonstrated in various in vivo and in vitro models, with effects on various modeled neuropathological processes. However, the therapeutic potential of macroautophagy-enhancing agents may be limited due to their lack of selectivity and the double-edged sword properties of macroautophagy. Recently developed compounds that selectively target downstream components of the ALP, including GCCase, TFEB and CMA elements, exert more specific effects on autophagy and may have exciting therapeutic perspective. Although several fundamental questions need to be further addressed before these novel agents can be applied in a clinical setting, the research field of autophagy is developing quickly and clinically relevant updates on these topics may be expected soon. Further characterization of dysfunctional autophagy in different stages as well as genetic and molecular subtypes of PD in combination with the effective clinical translation of downstream autophagy regulation offers exciting new avenues for the development of therapeutic strategies in PD.

Abbreviations

6-OHDA: 6-Hydroxydopamine; AICAR: Aminoimidazole-4-carboxamide ribonucleotide; ALP: Autophagy-lysosomal pathway; AMBRA1: Autophagy/Beclin-1 regulator 1; AMPK: AMP activated protein kinase; aSMase: Acid sphingomyelinase; ATP13A2: P-type ATPase; BH3 mimetics: Beclin-1 expression mimetics; CMA: Chaperone-mediated autophagy; DLB: Dementia with Lewy Bodies; ER: Endoplasmic reticulum; FCCP: Trifluorocarbonylcyanide phenylhydrazone; GBA1: Glucosidase beta acid 1; GCCase: Glucocerebrosidase; GD: Gaucher Disease; G α : Ca²⁺-calpain-G-stimulatory protein α ; Hsc70: Heat-shock cognate protein of 70 kDa; IMPase: Inositol monophosphatase; Ins: Inositol; IP₃: 1,4,5-Triphosphate (IP₃); LAMP: Lysosomal-associated membrane protein; LB: Lewy Bodies; LC3: Microtubule-associated protein 1 light chain 3; LIMP2: Lysosomal integral membrane protein type 2; *LRRK2*: Leucine-rich repeat kinase 2; LSD: Lysosomal storage disorder; miR: micro-RNA; MPTP: Methyl-4-phenyl-1,2,3,6 tetrahydropyridine; mTOR: Mammalian target of rapamycin; PD: Parkinson's Disease; PML1: p62-Mediated mitophagy inducer; PREP: Prolyl oligopeptidase; SCARB2: Scavenger receptor class B member 2; SMER: Small-molecule enhancer of rapamycin; *SMPD1*: Sphingomyelin phosphodiesterase 1; TFEB: Transcription factor EB; TSC1/TSC2 complex: Tuberous sclerosis complex; ULK-1: UNC-51-like kinase 1; UPS: Ubiquitin-proteasomal system; *VPS35*: Vacuolar protein sorting protein-associated protein 35; WT: Wild-type; α -syn: α -synuclein

Acknowledgements

Not applicable.

Funding

Not applicable (No relevant funding for the research reported).

Availability of data and materials

Not applicable.

Authors' contributions

TM proposed the writing of the review paper. TM and WvdB wrote and revised the manuscript. TM constructed the Figures. JH, AI, TB, LP and MC provided scientific input and were involved in the revision of the manuscript. All authors read and approved the final manuscript.

Competing interests

The authors declare that they have no competing interests.

Consent for publication

Not applicable.

Ethics approval and consent to participate

Not applicable.

Author details

¹Department of Anatomy and Neurosciences, Section Clinical Neuroanatomy, Amsterdam Neuroscience, VU University Medical Center Amsterdam, Amsterdam, The Netherlands. ²Department of Pathology, Amsterdam Neuroscience, VU University Medical Center Amsterdam, Amsterdam, The Netherlands. ³Department of Pharmaceutical Sciences, University of Perugia, Perugia, Italy. ⁴Department of Medicine, Section of Neurology, University of Perugia, Perugia, Italy. ⁵UMR-S 1172—JPArC—Centre de Recherche Jean-Pierre AUBERT Neurosciences et Cancer, University of Lille, Lille F-59000, France. ⁶Inserm, UMR-S 1172, Team "Early stages of Parkinson's disease", F-59000 Lille, France.

Received: 7 June 2016 Accepted: 18 January 2017

Published online: 25 January 2017

References

- Spillantini MG, Crowther RA, Jakes R, Hasegawa M, Goedert M. alpha-Synuclein in filamentous inclusions of Lewy bodies from Parkinson's disease and dementia with lewy bodies. *Proc Natl Acad Sci U S A*. 1998;95:6469–73.
- Xilouri M, Brekk OR, Stefanis L. alpha-Synuclein and protein degradation systems: a reciprocal relationship. *Mol Neurobiol*. 2013;47:537–51. doi:10.1007/s12035-012-8341-2.
- Ebrahimi-Fakhari D, Wahlster L, Mclean PJ. Protein degradation pathways in Parkinson's disease: curse or blessing. *Acta Neuropathol*. 2012;124:153–72. doi:10.1007/s00401-012-1004-6.
- Webb JL, Ravikumar B, Atkins J, Skepper JN, Rubinsztein DC. Alpha-Synuclein is degraded by both autophagy and the proteasome. *J Biol Chem*. 2003;278:25009–13.
- Xilouri M, Brekk OR, Stefanis L. Autophagy and alpha-synuclein: relevance to Parkinson's disease and related synucleopathies. *Mov Disord*. 2016;31:178–92. doi:10.1002/mds.26477.
- Hara T, Nakamura K, Matsui M, Yamamoto A, Nakahara Y, Suzuki-Migishima R, et al. Suppression of basal autophagy in neural cells causes neurodegenerative disease in mice. *Nature*. 2006;441:885–9.
- Komatsu M, Waguri S, Chiba T, Murata S, Iwata J, Tanida I, et al. Loss of autophagy in the central nervous system causes neurodegeneration in mice. *Nature*. 2006;441:880–4.
- Nixon RA. The role of autophagy in neurodegenerative disease. *Nat Med*. 2013;19:983–97. doi:10.1038/nm.3232.
- Lynch-Day MA, Mao K, Wang K, Zhao M, Klionsky DJ. The role of autophagy in Parkinson's disease. *Cold Spring Harb Perspect Med*. 2012;2:a009357. doi:10.1101/cshperspect.a009357.
- Bove J, Martinez-Vicente M, Vila M. Fighting neurodegeneration with rapamycin: mechanistic insights. *Nat Rev Neurosci*. 2011;12:437–52. doi:10.1038/nrn3068.
- Rubinsztein DC, Codogno P, Levine B. Autophagy modulation as a potential therapeutic target for diverse diseases. *Nat Rev Drug Discov*. 2012;11:709–30. doi:10.1038/nrd3802.
- Li WW, Li J, Bao JK. Microautophagy: lesser-known self-eating. *Cell Mol Life Sci*. 2012;69:1125–36. doi:10.1007/s00018-011-0865-5.
- Ravikumar B, Sarkar S, Davies JE, Futter M, Garcia-Arencibia M, Green-Thompson ZW, et al. Regulation of mammalian autophagy in physiology and pathophysiology. *Physiol Rev*. 2010;90:1383–435. doi:10.1152/physrev.00030.2009.
- Cuervo AM. Autophagy: in sickness and in health. *Trends Cell Biol*. 2004;14:70–7.
- Reggiori F, Komatsu M, Finley K, Simonsen A. Autophagy: more than a nonselective pathway. *Int J Cell Biol*. 2012;2012:219625. doi:10.1155/2012/219625.
- Farré JC, Subramani S. Mechanistic insights into selective autophagy pathways: lessons from yeast. *Nat Rev Mol Cell Biol*. 2016;17(9):537–52. doi:10.1038/nrm.2016.74.
- Khaminets A, Behl C, Dikic I. Ubiquitin-dependent and independent signals in selective autophagy. *Trends Cell Biol*. 2016;26(1):6–16. doi:10.1016/j.tcb.2015.08.010.
- Bento CF, Renna M, Ghislat G, Puri C, Ashkenazi A, Vicinanza M, et al. Mammalian autophagy: How does it work? *Annu Rev Biochem*. 2016;85:685–713. doi:10.1146/annurev-biochem-060815-014556.
- Laplante M, Sabatini DM. mTOR signaling in growth control and disease. *Cell*. 2012;149:274–93. doi:10.1016/j.cell.2012.03.017.
- Kim J, Kundu M, Viollet B, Guan KL. AMPK and mTOR regulate autophagy through direct phosphorylation of Ulk1. *Nat Cell Biol*. 2011;13:132–41. doi:10.1038/ncb2152.
- Gao X, Zhang Y, Arrazola P, Hino O, Kobayashi T, Yeung RS, et al. Tsc tumour suppressor proteins antagonize amino-acid-TOR signalling. *Nat Cell Biol*. 2002;4:699–704.
- Itakura E, Kishi C, Inoue K, Mizushima N. Beclin 1 forms two distinct phosphatidylinositol 3-kinase complexes with mammalian Atg14 and UVRAG. *Mol Biol Cell*. 2008;19:5360–72. doi:10.1091/mbc.E08-01-0080.
- Choi AM, Ryter SW, Levine B. Autophagy in human health and disease. *N Engl J Med*. 2013;368:1845–6. doi:10.1056/NEJMr1205406.
- Alers S, Löffler AS, Wesselborg S, Stork B. Role of AMPK-mTOR-Ulk1/2 in the regulation of autophagy: cross talk, shortcuts, and feedbacks. *Mol Cell Biol*. 2012;32(1):2–11. doi:10.1128/MCB.06159-11.
- Sardiello M, Palmieri M, di Ronza A, Medina DL, Valenza M, Gennarino VA, et al. A gene network regulating lysosomal biogenesis and function. *Science*. 2009;325:473–7. doi:10.1126/science.1174447.
- Settembre C, Di MC, Polito VA, Garcia AM, Vetrini F, Erdin S, et al. TFEB links autophagy to lysosomal biogenesis. *Science*. 2011;332:1429–33. doi:10.1126/science.1204592.
- Settembre C, Zoncu R, Medina DL, Vetrini F, Erdin S, Erdin S, et al. A lysosome-to-nucleus signalling mechanism senses and regulates the lysosome via mTOR and TFEB. *EMBO J*. 2012;31:1095–108. doi:10.1038/emboj.2012.32.
- Zoncu R, Bar-Peled L, Efeyan A, Wang S, Sancak Y, Sabatini DM. mTORC1 senses lysosomal amino acids through an inside-out mechanism that requires the vacuolar H(+)-ATPase. *Science*. 2011;334:678–83. doi:10.1126/science.1207056.
- Williams A, Sarkar S, Cuddon P, Tfofi EK, Saiki S, Siddiqi FH, et al. Novel targets for Huntington's disease in an mTOR-independent autophagy pathway. *Nat Chem Biol*. 2008;4:295–305. doi:10.1038/nchembio.79.
- Criollo A, Maiuri MC, Tasdemir E, Vitale I, Fiebig AA, Andrews D, et al. Regulation of autophagy by the inositol trisphosphate receptor. *Cell Death Differ*. 2007;14:1029–39.
- Gan-Or Z, Dion PA, Rouleau GA. Genetic perspective on the role of the autophagy-lysosome pathway in Parkinson disease. *Autophagy*. 2015;11:1443–57. doi:10.1080/15548627.2015.1067364.
- Sidransky E, Lopez G. The link between the GBA gene and parkinsonism. *Lancet Neurol*. 2012;11:986–98. doi:10.1080/15548627.2015.1067364.
- Sidransky E, Nalls MA, Aasly JO, Aharon-Peretz J, Annesi G, Barbosa ER, et al. Multicenter analysis of glucocerebrosidase mutations in Parkinson's disease. *N Engl J Med*. 2009;361:1651–61. doi:10.1056/NEJMoa0901281.
- Liu J, Zhang HX. Significant study of population stratification, sensitivity analysis and trim and fill analyses on GBA mutation and Parkinson's disease. *Am J Med Genet B Neuropsychiatr Genet*. 2014;165B:96–102. doi:10.1002/ajmg.b.32214.
- Mao X, Wang T, Peng R, Chang X, Li N, Gu Y, et al. Mutations in GBA and risk of Parkinson's disease: a meta-analysis based on 25 case-control studies. *Neurol Res*. 2013;35:873–8. doi:10.1179/1743132813Y.0000000224.
- Schapira AH. Glucocerebrosidase and Parkinson disease: recent advances. *Mol Cell Neurosci*. 2015;66:37–42. doi:10.1016/j.mcn.2015.03.013.

37. Moors T, Paciotti S, Chiasserini D, Calabresi P, Parnetti L, Beccari T, et al. Lysosomal Dysfunction and alpha-Synuclein Aggregation in Parkinson's Disease: Diagnostic Links. *Mov Disord* 2016. doi:10.1002/mds.26562.
38. Reczek D, Schwake M, Schroder J, Hughes H, Blanz J, Jin X, et al. LIMP-2 is a receptor for lysosomal mannose-6-phosphate-independent targeting of beta-glucocerebrosidase. *Cell*. 2007;131:770–83.
39. Dagan E, Schlesinger I, Ayoub M, Mory A, Nassar M, Kurolop A, et al. The contribution of niemann-pick SMPD1 mutations to Parkinson disease in Ashkenazi Jews. *Parkinsonism Relat Disord*. 2015;21:1067–71. doi:10.1016/j.parkreldis.2015.06.016.
40. Foo JN, Liany H, Bei JX, Yu XQ, Liu J, Au WL, et al. Rare lysosomal enzyme gene SMPD1 variant (p. R591C) associates with Parkinson's disease. *Neurobiol Aging*. 2013;34:2890–5. doi:10.1016/j.neurobiolaging.2013.06.010.
41. Gan-Or Z, Ozelius LJ, Bar-Shira A, Saunders-Pullman R, Mirelman A, Kornreich R, et al. The p. L302P mutation in the lysosomal enzyme gene SMPD1 is a risk factor for Parkinson disease. *Neurology*. 2013;80:1606–10. doi:10.1212/WNL.0b013e31828f180e.
42. Wu RM, Lin CH. The p.L302P mutation in the lysosomal enzyme gene SMPD1 is a risk factor for Parkinson disease. *Neurology*. 2014;82:283. doi:10.1212/WNL.0000000000000004.
43. Trinh J, Farrer M. Advances in the genetics of Parkinson disease. *Nat Rev Neurol*. 2013;9:445–54. doi:10.1038/nrneurol.2013.132.
44. Burchell VS, Nelson DE, Sanchez-Martinez A, Delgado-Camprubi M, Ivatt RM, Pogson JH, et al. The Parkinson's disease-linked proteins Fbxo7 and Parkin interact to mediate mitophagy. *Nat Neurosci*. 2013;16:1257–65. doi:10.1038/nn.3489.
45. Orenstein SJ, Kuo SH, Tasset I, Arias E, Koga H, Fernandez-Carasa I, et al. Interplay of LRRK2 with chaperone-mediated autophagy. *Nat Neurosci*. 2013;16:394–406. doi:10.1038/nn.3350.
46. Park S, Han S, Choi I, Kim B, Park SP, Joe EH, et al. Interplay between leucine-rich repeat kinase 2 (LRRK2) and p62/SQSTM-1 in selective autophagy. *Plos One*. 2016;11:e0163029. doi:10.1371/journal.pone.0163029.
47. Plowey ED, Cherra III SJ, Liu YJ, Chu CT. Role of autophagy in G2019S-LRRK2-associated neurite shortening in differentiated SH-SY5Y cells. *J Neurochem*. 2008;105:1048–56. doi:10.1111/j.1471-4159.2008.05217.
48. Anglade P, Vyas S, Javoy-Agid F, Herrero MT, Michel PP, Marquez J, et al. Apoptosis and autophagy in nigral neurons of patients with Parkinson's disease. *Histol Histopathol*. 1997;12:25–31.
49. Toulorge D, Schapira AH, Hajj R. Molecular changes in the postmortem parkinsonian brain. *J Neurochem*. 2016;139 Suppl 1:27–58. doi:10.1111/jnc.13696.
50. Dehay B, Bove J, Rodriguez-Muela N, Perier C, Recasens A, Boya P, et al. Pathogenic lysosomal depletion in Parkinson's disease. *J Neurosci*. 2010;30:12535–44. doi:10.1523/JNEUROSCI.1920-10.2010.
51. Tanji K, Mori F, Kakita A, Takahashi H, Wakabayashi K. Alteration of autophagosomal proteins (LC3, GABARAP and GATE-16) in Lewy body disease. *Neurobiol Dis*. 2011;43:690–7. doi:10.1016/j.nbd.2011.05.022.
52. Chu Y, Dodiya H, Aebischer P, Olanow CW, Kordower JH. Alterations in lysosomal and proteasomal markers in Parkinson's disease: relationship to alpha-synuclein inclusions. *Neurobiol Dis*. 2009;35:385–98. doi:10.1016/j.nbd.2009.05.023.
53. Alvarez-Erviti L, Rodriguez-Oroz MC, Cooper JM, Caballero C, Ferrer I, Obeso JA, et al. Chaperone-mediated autophagy markers in Parkinson disease brains. *Arch Neurol*. 2010;67:1464–72. doi:10.1001/archneurol.2010.198.
54. Murphy KE, Gysbers AM, Abbott SK, Spiro AS, Furuta A, Cooper A, et al. Lysosomal-associated membrane protein 2 isoforms are differentially affected in early Parkinson's disease. *Mov Disord*. 2015;30(12):1639–47. doi:10.1002/mds.26141.
55. Gegg ME, Burke D, Heales SJ, Cooper JM, Hardy J, Wood NW, et al. Glucocerebrosidase deficiency in substantia nigra of parkinson disease brains. *Ann Neurol*. 2012;72:455–63. doi:10.1002/ana.23614.
56. Murphy KE, Gysbers AM, Abbott SK, Tayebi N, Kim WS, Sidransky E, et al. Reduced glucocerebrosidase is associated with increased alpha-synuclein in sporadic Parkinson's disease. *Brain*. 2014;137:834–48. doi:10.1093/brain/awt367.
57. Chiasserini D, Paciotti S, Eusebi P, Persichetti E, Tasegian A, Kurzawa-Akanbi M, et al. Selective loss of glucocerebrosidase activity in sporadic Parkinson's disease and dementia with Lewy bodies. *Mol Neurodegener*. 2015;10:15. doi:10.1186/s13024-015-0010-2.
58. Balducci C, Pierguidi L, Persichetti E, Parnetti L, Sbaragli M, Tassi C, et al. Lysosomal hydrolases in cerebrospinal fluid from subjects with Parkinson's disease. *Mov Disord*. 2007;22:1481–4.
59. van Dijk KD, Persichetti E, Chiasserini D, Eusebi P, Beccari T, Calabresi P, et al. Changes in endolysosomal enzyme activities in cerebrospinal fluid of patients with Parkinson's disease. *Mov Disord*. 2013;28:747–54. doi:10.1002/mds.25495.
60. Parnetti L, Chiasserini D, Persichetti E, Eusebi P, Varghese S, Qureshi MM, et al. Cerebrospinal fluid lysosomal enzymes and alpha-synuclein in Parkinson's disease. *Mov Disord*. 2014;29:1019–27. doi:10.1002/mds.25772.
61. Rothaug M, Zunke F, Mazzulli JR, Schweizer M, Altmepfen H, Lullmann-Rauch R, et al. LIMP-2 expression is critical for beta-glucocerebrosidase activity and alpha-synuclein clearance. *Proc Natl Acad Sci U S A*. 2014;111:15573–8. doi:10.1073/pnas.1405700111.
62. Mantle D, Falkous G, Ishiura S, Perry RH, Perry EK. Comparison of cathepsin protease activities in brain tissue from normal cases and cases with Alzheimer's disease, Lewy body dementia, Parkinson's disease and Huntington's disease. *J Neurol Sci*. 1995;131:65–70.
63. Murphy KE, Cottle L, Gysbers AM, Cooper AA, Halliday GM. ATP13A2 (PARK9) protein levels are reduced in brain tissue of cases with Lewy bodies. *Acta Neuropathol Commun*. 2013;1:11. doi:10.1186/2051-5960-1-11.
64. Ramonet D, Podhajska A, Stafa K, Sonnay S, Trancikova A, Tsika E, et al. PARK9-associated ATP13A2 localizes to intracellular acidic vesicles and regulates cation homeostasis and neuronal integrity. *Hum Mol Genet*. 2012;21:1725–43. doi:10.1093/hmg/ddr606.
65. Dijkstra AA, Ingrassia A, de Menezes RX, van Kesteren RE, Rozemuller AJ, Heutink P, et al. Evidence for immune response, axonal dysfunction and reduced endocytosis in the substantia nigra in early stage Parkinson's disease. *Plos One*. 2015;10:e0128651. doi:10.1371/journal.pone.0128651.
66. Elstner M, Morris CM, Heim K, Bender A, Mehta D, Jaros E, et al. Expression analysis of dopaminergic neurons in Parkinson's disease and aging links transcriptional dysregulation of energy metabolism to cell death. *Acta Neuropathol*. 2011;122:75–86. doi:10.1007/s00401-011-0828-9.
67. Mutez E, Nkiliza A, Belarbi K, de Broucker A, Vanbesien-Mailliot C, Bleuse S, et al. Involvement of the immune system, endocytosis and EIF2 signaling in both genetically determined and sporadic forms of Parkinson's disease. *Neurobiol Dis*. 2014;63:165–70. doi:10.1016/j.nbd.2013.11.007.
68. Crews L, Spencer B, Desplats P, Patrick C, Paulino A, Rockenstein E, et al. Selective molecular alterations in the autophagy pathway in patients with Lewy body disease and in models of alpha-synucleinopathy. *Plos One*. 2010;5:e9313. doi:10.1371/journal.pone.0009313.
69. Miki Y, Tanji K, Mori F, Utsumi J, Sasaki H, Kakita A, et al. Alteration of upstream autophagy-related proteins (ULK1, ULK2, Beclin1, VPS34 and AMBRA1) in Lewy body disease. *Brain Pathol*. 2015;26(3):359–70. doi:10.1111/bpa.12297.
70. Decressac M, Mattsson B, Weikop P, Lundblad M, Jakobsson J, Bjorklund A. TFEB-mediated autophagy rescues midbrain dopamine neurons from alpha-synuclein toxicity. *Proc Natl Acad Sci U S A*. 2013;110:E1817–26. doi:10.1073/pnas.1305623110.
71. Vogiatzi T, Xilouri M, Vekrellis K, Stefanis L. Wild type alpha-synuclein is degraded by chaperone-mediated autophagy and macroautophagy in neuronal cells. *J Biol Chem*. 2008;283:23542–56. doi:10.1074/jbc.M801992200.
72. Yu WH, Dorado B, Figueroa HY, Wang L, Planel E, Cookson MR, et al. Metabolic activity determines efficacy of macroautophagic clearance of pathological oligomeric alpha-synuclein. *Am J Pathol*. 2009;175:736–47. doi:10.2353/ajpath.2009.080928.
73. Cuervo AM, Stefanis L, Fredenburg R, Lansbury PT, Sulzer D. Impaired degradation of mutant alpha-synuclein by chaperone-mediated autophagy. *Science*. 2004;305:1292–5.
74. Oueslati A, Schneider BL, Aebischer P, Lashuel HA. Polo-like kinase 2 regulates selective autophagic alpha-synuclein clearance and suppresses its toxicity in vivo. *Proc Natl Acad Sci U S A*. 2013;110:E3945–54. doi:10.1073/pnas.1309991110.
75. Tenreiro S, Reimao-Pinto MM, Antas P, Rino J, Wawrzycka D, Macedo D, et al. Phosphorylation modulates clearance of alpha-synuclein inclusions in a yeast model of Parkinson's disease. *Plos Genet*. 2014;10:e1004302. doi:10.1371/journal.pgen.1004302.
76. Mazzulli JR, Xu YH, Sun Y, Knight AL, Mclean PJ, Caldwell GA, et al. Gaucher disease glucocerebrosidase and alpha-synuclein form a bidirectional pathogenic loop in synucleinopathies. *Cell*. 2011;146:37–52. doi:10.1016/j.cell.2011.06.001.
77. Manning-Bog AB, Schule B, Langston JW. Alpha-synuclein-glucocerebrosidase interactions in pharmacological gaucher models: a biological link between gaucher disease and parkinsonism. *Neurotoxicology*. 2009;30:1127–32. doi:10.1016/j.neuro.2009.06.009.

78. Sardi SP, Clarke J, Kinnecom C, Tamsett TJ, Li L, Stanek LM, et al. CNS expression of glucocerebrosidase corrects alpha-synuclein pathology and memory in a mouse model of gaucher-related synucleinopathy. *Proc Natl Acad Sci U S A*. 2011;108:12101–6. doi:10.1073/pnas.1108197108.
79. Bae EJ, Yang NY, Lee C, Kim S, Lee HJ, Lee SJ. Haploinsufficiency of cathepsin D leads to lysosomal dysfunction and promotes cell-to-cell transmission of alpha-synuclein aggregates. *Cell Death Dis*. 2015;6:e1901. doi:10.1038/cddis.2015.283.
80. Bae EJ, Yang NY, Song M, Lee CS, Lee JS, Jung BC, et al. Glucocerebrosidase depletion enhances cell-to-cell transmission of alpha-synuclein. *Nat Commun*. 2014;5:4755. doi:10.1038/ncomms5755.
81. Maiese K, Chong ZZ, Shang YC, Wang S: mTOR: on target for novel therapeutic strategies in the nervous system. *Trends Mol Med* 2013, 19: 51–60. doi:10.1016/j.molmed.2012.11.001. DOI: 10.1016/j.neuroscience.2009.08.014.
82. Pan T, Rawal P, Wu Y, Xie W, Jankovic J, Le W. Rapamycin protects against rotenone-induced apoptosis through autophagy induction. *Neuroscience*. 2009;164:541–51. doi:10.1016/j.neuroscience.2009.08.014.
83. Malagelada C, Jin ZH, Jackson-Lewis V, Przedborski S, Greene LA. Rapamycin protects against neuron death in in vitro and in vivo models of Parkinson's disease. *J Neurosci*. 2010;30:1166–75. doi:10.1523/JNEUROSCI.3944-09.2010.
84. Bai X, Wey MC, Fernandez E, Hart MJ, Gelfond J, Bokov AF, et al. Rapamycin improves motor function, reduces 4-hydroxynonenal adducted protein in brain, and attenuates synaptic injury in a mouse model of synucleinopathy. *Pathobiol Aging Age Relat Dis*. 2015;5:28743. doi:10.3402/pba.v5.28743.
85. Santini E, Heiman M, Greengard P, Valjent E, Fisone G. Inhibition of mtor signaling in Parkinson's disease prevents L-DOPA-induced dyskinesia. *Sci Signal*. 2009;2:ra36. doi:10.1126/scisignal.2000308.
86. Decressac M, Bjorklund A. mTOR inhibition alleviates L-DOPA-induced dyskinesia in parkinsonian rats. *J Parkinsons Dis*. 2013;3:13–7. doi:10.3233/JPD-120155.
87. Tain LS, Mortiboys H, Tao RN, Ziviani E, Bandmann O, Whitworth AJ. Rapamycin activation of 4E-BP prevents parkinsonian dopaminergic neuron loss. *Nat Neurosci*. 2009;12:1129–35. doi:10.1038/nn.2372.
88. Sarkar S, Floto RA, Berger Z, Imarisio S, Cordenier A, Pasco M, et al. Lithium induces autophagy by inhibiting inositol monophosphatase. *J Cell Biol*. 2005;170:1101–11.
89. Forlenza OV, De-Paula VJ, Diniz BS. Neuroprotective effects of lithium: implications for the treatment of Alzheimer's disease and related neurodegenerative disorders. *ACS Chem Neurosci*. 2014;5:443–50. doi:10.1021/cn5000309.
90. Lazzara CA, Kim YH. Potential application of lithium in Parkinson's and other neurodegenerative diseases. *Front Neurosci*. 2015;9:403. doi:10.3389/fnins.2015.00403.
91. Oruch R, Elderbi MA, Khattab HA, Pryme IF, Lund A. Lithium: a review of pharmacology, clinical uses, and toxicity. *Eur J Pharmacol*. 2014;740C:464–73. doi:10.1016/j.ejphar.2014.06.042.
92. Hou L, Xiong N, Liu L, Huang J, Han C, Zhang G, et al. Lithium protects dopaminergic cells from rotenone toxicity via autophagy enhancement. *BMC Neurosci*. 2015;16:82. doi:10.1186/s12868-015-0222-y.
93. Xiong N, Jia M, Chen C, Xiong J, Zhang Z, Huang J, et al. Potential autophagy enhancers attenuate rotenone-induced toxicity in SH-SY5Y. *Neuroscience*. 2011;199:292–302. doi:10.1016/j.neuroscience.2011.10.031.
94. Li XZ, Chen XP, Zhao K, Bai LM, Zhang H, Zhou XP. Therapeutic effects of valproate combined with lithium carbonate on MPTP-induced parkinsonism in mice: possible mediation through enhanced autophagy. *Int J Neurosci*. 2013;123:73–9. doi:10.3109/00207454.2012.729234.
95. Ng CH, Guan MS, Koh C, Ouyang X, Yu F, Tan EK, et al. AMP kinase activation mitigates dopaminergic dysfunction and mitochondrial abnormalities in *Drosophila* models of Parkinson's disease. *J Neurosci*. 2012;32:14311–7. doi:10.1523/JNEUROSCI.0499-12.2012.
96. Patil SP, Jain PD, Ghumatkar PJ, Tambe R, Sathaye S. Neuroprotective effect of metformin in MPTP-induced Parkinson's disease in mice. *Neuroscience*. 2014;26(277):747–54. doi:10.1016/j.neuroscience.2014.07.046.
97. Dulovic M, Jovanovic M, Xilouri M, Stefanis L, Harhaji-Trajkovic L, Kravic-Stevovic T, et al. The protective role of AMP-activated protein kinase in alpha-synuclein neurotoxicity in vitro. *Neurobiol Dis*. 2014;63:1–11. doi:10.1016/j.nbd.2013.11.002.
98. Perez-Revuelta BI, Hettich MM, Ciociaro A, Rotermund C, Kahle PJ, Krauss S, et al. Metformin lowers Ser-129 phosphorylated alpha-synuclein levels via mtor-dependent protein phosphatase 2A activation. *Cell Death Dis*. 2014;5:e1209. doi:10.1038/cddis.2014.175.
99. Ur Rasheed MS, Tripathi MK, Mishra AK, Shukla S, Singh MP. Resveratrol protects from toxin-induced parkinsonism: plethora of proofs hitherto petty translational value. *Mol Neurobiol*. 2015;53(5):2751–60. doi:10.1007/s12035-015-9124-3.
100. Wu Y, Li X, Zhu JX, Xie W, Le W, Fan Z, et al. Resveratrol-activated AMPK/SIRT1/autophagy in cellular models of Parkinson's disease. *Neurosignals*. 2011;19:163–74. doi:10.1159/000328516.
101. Lin TK, Chen SD, Chuang YC, Lin HY, Huang CR, Chuang JH, et al. Resveratrol partially prevents rotenone-induced neurotoxicity in dopaminergic SH-SY5Y cells through induction of heme oxygenase-1 dependent autophagy. *Int J Mol Sci*. 2014;15:1625–46. doi:10.3390/ijms15011625.
102. Ferretta A, Gaballo A, Tanzarella P, Piccoli C, Capitanio N, Nico B, et al. Effect of resveratrol on mitochondrial function: implications in parkin-associated familial Parkinson's disease. *Biochim Biophys Acta*. 2014;1842:902–15. doi:10.1016/j.bbdis.2014.02.010.
103. De Bosch BJ, Heitmeier MR, Mayer AL, Higgins CB, Crowley JR, Kraft TE, et al. Trehalose inhibits solute carrier 2A (SLC2A) proteins to induce autophagy and prevent hepatic steatosis. *Sci Signal*. 2016;9:ra21.
104. Sarkar S, Davies JE, Huang Z, Tunnacliffe A, Rubinsztein DC. Trehalose, a novel mTOR-independent autophagy enhancer, accelerates the clearance of mutant huntingtin and alpha-synuclein. *J Biol Chem*. 2007;282:5641–52.
105. Casarejos MJ, Solano RM, Gomez A, Perucho J, de Yébenes JG, Mena MA. The accumulation of neurotoxic proteins, induced by proteasome inhibition, is reverted by trehalose, an enhancer of autophagy, in human neuroblastoma cells. *Neurochem Int*. 2011;58:512–20. doi:10.1016/j.neuint.2011.01.008.
106. Lan DM, Liu FT, Zhao J, Chen Y, Wu JJ, Ding ZT, et al. Effect of trehalose on PC12 cells overexpressing wild-type or A53T mutant alpha-synuclein. *Neurochem Res*. 2012;37:2025–32. doi:10.1007/s11064-012-0823-0.
107. Wu F, Xu HD, Guan JJ, Hou YS, Gu JH, Zhen XC, et al. Rotenone impairs autophagic flux and lysosomal functions in Parkinson's disease. *Neuroscience*. 2015;284:900–11. doi:10.1016/j.neuroscience.2014.11.004.
108. Sarkar S, Chigurupati S, Raymick J, Mann D, Bowyer JF, Schmitt T, et al. Neuroprotective effect of the chemical chaperone, trehalose in a chronic MPTP-induced Parkinson's disease mouse model. *Neurotoxicology*. 2014;44C:250–62. doi:10.1016/j.neuro.2014.07.006.
109. He Q, Koprach JB, Wang Y, Yu WB, Xiao BG, Brotchie JM, et al. Treatment with trehalose prevents behavioral and neurochemical deficits produced in an AAV alpha-synuclein rat model of Parkinson's disease. *Mol Neurobiol*. 2015;53(4):2258–68. doi:10.1007/s12035-015-9173-7.
110. Tanji K, Miki Y, Maruyama A, Mimura J, Matsumiya T, Mori F, et al. Trehalose intake induces chaperone molecules along with autophagy in a mouse model of Lewy body disease. *Biochem Biophys Res Commun*. 2015;465:746–52. doi:10.1016/j.bbrc.2015.08.076.
111. Lee GC, Lin CH, Tao YC, Yang JM, Hsu KC, Huang YJ, et al. The potential of lactulose and melibiose, two novel trehalase-indigestible and autophagy-inducing disaccharides, for polyQ-mediated neurodegenerative disease treatment. *Neurotoxicology*. 2015;48:120–30. doi:10.1016/j.neuro.2015.03.009.
112. Sarkar S, Perlstein EO, Imarisio S, Pineau S, Cordenier A, Maglathlin RL, et al. Small molecules enhance autophagy and reduce toxicity in Huntington's disease models. *Nat Chem Biol*. 2007;3:331–8.
113. Sarkar S, Rubinsztein DC. Small molecule enhancers of autophagy for neurodegenerative diseases. *Mol Biosyst*. 2008;4:895–901. doi:10.1039/b804606a.
114. Bharadwaj PR, Verdile G, Barr RK, Gupta V, Steele JW, Lachenmayer ML, et al. Latrepirdine (dimebon) enhances autophagy and reduces intracellular GFP-Abeta42 levels in yeast. *J Alzheimers Dis*. 2012;32:949–67. doi:10.3233/JAD-2012-120178.
115. Steele JW, Ju S, Lachenmayer ML, Liken J, Stock A, Kim SH, et al. Latrepirdine stimulates autophagy and reduces accumulation of alpha-synuclein in cells and in mouse brain. *Mol Psychiatry*. 2013;18:882–8. doi:10.1038/mp.2012.115.
116. Bharadwaj PR, Bates KA, Porter T, Teimouri E, Perry G, Steele JW, et al. Latrepirdine: molecular mechanisms underlying potential therapeutic roles in Alzheimer's and other neurodegenerative diseases. *Transl Psychiatry*. 2013;3:e332. doi:10.1038/tp.2013.97.
117. Olanow CW, Schapira AH. Therapeutic prospects for Parkinson disease. *Ann Neurol*. 2013;74:337–47. doi:10.1002/ana.24011.
118. Buttner S, Broeskamp F, Sommer C, Markaki M, Habernig L, Alavian-Ghavanini A, et al. Spermidine protects against alpha-synuclein neurotoxicity. *Cell Cycle*. 2014;13:3903–8. doi:10.4161/15384101.2014.973309.
119. Jiang TF, Zhang YJ, Zhou HY, Wang HM, Tian LP, Liu J, et al. Curcumin ameliorates the neurodegenerative pathology in A53T alpha-synuclein cell

- model of Parkinson's disease through the downregulation of mTOR/p70S6K signaling and the recovery of macroautophagy. *J Neuroimmune Pharmacol*. 2013;8:356–69. doi:10.1007/s11481-012-9431-7.
120. Filomeni G, Graziani I, De ZD, Dini L, Centonze D, Rotilio G, et al. Neuroprotection of kaempferol by autophagy in models of rotenone-mediated acute toxicity: possible implications for Parkinson's disease. *Neurobiol Aging*. 2012;33:767–85. doi:10.1016/j.neurobiolaging.2010.05.021.
 121. Macedo D, Tavares L, McDougall GJ, Vicente MH, Stewart D, Ferreira RB, et al. (Poly)phenols protect from alpha-synuclein toxicity by reducing oxidative stress and promoting autophagy. *Hum Mol Genet*. 2015;24:1717–32. doi:10.1093/hmg/ddu585.
 122. Hebrun ML, Lonskaya I, Moussa CE. Nilotinib reverses loss of dopamine neurons and improves motor behavior via autophagic degradation of alpha-synuclein in Parkinson's disease models. *Hum Mol Genet*. 2013;22:3315–28. doi:10.1093/hmg/ddt192.
 123. Mahul-Mellier AL, Fauvet B, Gysbers A, Dikiy I, Oueslati A, Georgeon S, et al. c-Abl phosphorylates alpha-synuclein and regulates its degradation: implication for alpha-synuclein clearance and contribution to the pathogenesis of Parkinson's disease. *Hum Mol Genet*. 2014;23:2858–79. doi:10.1093/hmg/ddt674.
 124. Rubinstein AD, Kimchi A. Life in the balance - a mechanistic view of the crosstalk between autophagy and apoptosis. *J Cell Sci*. 2012;125:5259–68. doi:10.1242/jcs.115865.
 125. Amaravadi R, Kimmelman AC, White E. Recent insights into the function of autophagy in cancer. *Genes Dev*. 2016;30:1913–30. doi:10.1101/gad.287524.116.
 126. White E. Deconvoluting the context-dependent role for autophagy in cancer. *Nat Rev Cancer*. 2012;12:401–10. doi:10.1038/nrc3262.
 127. Gomez-Santos C, Ferrer I, Santidrian AF, Barrachina M, Gil J, Ambrosio S. Dopamine induces autophagic cell death and alpha-synuclein increase in human neuroblastoma SH-SY5Y cells. *J Neurosci Res*. 2003;73:341–50.
 128. Stefanis L, Larsen KE, Rideout HJ, Sulzer D, Greene LA. Expression of A53T mutant but not wild-type alpha-synuclein in PC12 cells induces alterations of the ubiquitin-dependent degradation system, loss of dopamine release, and autophagic cell death. *J Neurosci*. 2001;21:9549–60.
 129. Choi KC, Kim SH, Ha JY, Kim ST, Son JH. A novel mTOR activating protein protects dopamine neurons against oxidative stress by repressing autophagy related cell death. *J Neurochem*. 2010;112:366–76. doi:10.1111/j.1471-4159.2009.06463.
 130. Xu Y, Liu C, Chen S, Ye Y, Guo M, Ren Q, et al. Activation of AMPK and inactivation of Akt result in suppression of mTOR-mediated S6K1 and 4E-BP1 pathways leading to neuronal cell death in *in vitro* models of Parkinson's disease. *Cell Signal*. 2014;26:1680–9. doi:10.1016/j.cellsig.2014.04.009.
 131. Cheng HC, Kim SR, Oo TF, Kareva T, Yarygina O, Rzhetskaya M, et al. Akt suppresses retrograde degeneration of dopaminergic axons by inhibition of macroautophagy. *J Neurosci*. 2011;31:2125–35. doi:10.1523/JNEUROSCI.5519-10.2011.
 132. Yang Y, Fukui K, Koike T, Zheng X. Induction of autophagy in neurite degeneration of mouse superior cervical ganglion neurons. *Eur J Neurosci*. 2007;26(10):2979–88. doi:10.1111/j.1460-9568.2007.05914.x.
 133. Xilouri M, Vogiatzi T, Vekrellis K, Park D, Stefanis L. Abberant alpha-synuclein confers toxicity to neurons in part through inhibition of chaperone-mediated autophagy. *Plos One*. 2009;4:e5515. doi:10.1371/journal.pone.0005515.
 134. Choubey V, Safulina D, Vaarmann A, Cagalinec M, Wareski P, Kuem M, et al. Mutant A53T alpha-synuclein induces neuronal death by increasing mitochondrial autophagy. *J Biol Chem*. 2011;286:10814–24. doi:10.1074/jbc.M110.132514.
 135. Liu Y, Levine B. Autophagy and autophagic cell death: the dark side of autophagy. *Cell Death Differ*. 2015;22:367–76. doi:10.1038/cdd.2014.143.
 136. Button RW, Luo S, Rubinsztein DC. Autophagic activity in neuronal cell death. *Neurosci Bull*. 2015;31:382–94. doi:10.1007/s12264-015-1528-y.
 137. Spencer B, Potkar R, Trejo M, Rockenstein E, Patrick C, Gindi R, et al. Beclin 1 gene transfer activates autophagy and ameliorates the neurodegenerative pathology in alpha-synuclein models of Parkinson's and Lewy body diseases. *J Neurosci*. 2009;29:13578–88. doi:10.1523/JNEUROSCI.4390-09.2009.
 138. Wang K, Huang J, Xie W, Huang L, Zhong C, Chen Z. Beclin1 and HMGB1 ameliorate the alpha-synuclein-mediated autophagy inhibition in PC12 cells. *Diagn Pathol*. 2016;11:15. doi:10.1186/s13000-016-0459-5.
 139. Obata T, Kubota S. Protective effect of tamoxifen on 1-methyl-4-phenylpyridine-induced hydroxyl radical generation in the rat striatum. *Neurosci Lett*. 2001;308:87–90.
 140. Savolainen MH, Richie CT, Harvey BK, Mannisto PT, Maguire-Zeiss KA, Myohanen TT. The beneficial effect of a prolyl oligopeptidase inhibitor, KYP-2047, on alpha-synuclein clearance and autophagy in A30P transgenic mouse. *Neurobiol Dis*. 2014;68:1–15. doi:10.1016/j.nbd.2014.04.003.
 141. Lu JH, Tan JQ, Durairajan SS, Liu LF, Zhang ZH, Ma L, et al. Isorhynchophylline, a natural alkaloid, promotes the degradation of alpha-synuclein in neuronal cells via inducing autophagy. *Autophagy*. 2012;8:98–108. doi:10.4161/auto.8.1.18313.
 142. Song W, Wang F, Lotfi P, Sardiello M, Segatori L. 2-Hydroxypropyl-beta-cyclodextrin promotes transcription factor EB-mediated activation of autophagy: implications for therapy. *J Biol Chem*. 2014;289:10211–22. doi:10.1074/jbc.M113.506246.
 143. Kilpatrick K, Zeng Y, Hancock T, Segatori L. Genetic and chemical activation of TFEB mediates clearance of aggregated alpha-synuclein. *Plos One*. 2015;10:e0120819. doi:10.1371/journal.pone.0120819.
 144. Baltazar GC, Guha S, Lu W, Lim J, Boesze-Battaglia K, Laties AM, et al. Acidic nanoparticles are trafficked to lysosomes and restore an acidic lysosomal pH and degradative function to compromised ARPE-19 cells. *Plos One*. 2012;7:e49635. doi:10.1371/journal.pone.0049635.
 145. Bourdenx M, Daniel J, Genin E, Soria FN, Blanchard-Desce M, Bezard E, et al. Nanoparticles restore lysosomal acidification defects: Implication for Parkinson and other lysosomal-related diseases. *Autophagy*. 2016;12(3):472–83. doi:10.1080/15548627.2015.1136769.
 146. Sardi SP, Clarke J, Viel C, Chan M, Tamsett TJ, Treleaven CM, et al. Augmenting CNS glucocerebrosidase activity as a therapeutic strategy for parkinsonism and other Gaucher-related synucleinopathies. *Proc Natl Acad Sci U S A*. 2013;110:3537–42. doi:10.1073/pnas.1220464110.
 147. Rocha EM, Smith GA, Park E, Cao H, Brown E, Hayes MA, et al. Glucocerebrosidase gene therapy prevents alpha-synucleinopathy of midbrain dopamine neurons. *Neurobiol Dis*. 2015;82:495–503. doi:10.1016/j.nbd.2015.09.009.
 148. Schapira AH, Gegg ME. Glucocerebrosidase in the pathogenesis and treatment of Parkinson disease. *Proc Natl Acad Sci U S A*. 2013;110:3214–5. doi:10.1073/pnas.1300822110.
 149. Blanz J, Saftig P. Parkinson's disease: acid-glucocerebrosidase activity and alpha-synuclein clearance. *J Neurochem*. 2016. DOI: 10.1111/jnc.13517
 150. McNeill A, Magalhaes J, Shen C, Chau KY, Hughes D, Mehta A, et al. Amroxol improves lysosomal biochemistry in glucocerebrosidase mutation-linked Parkinson disease cells. *Brain*. 2014;137:1481–95. doi:10.1093/brain/awu020.
 151. Ambrosi G, Ghezzi C, Zangaglia R, Levandis G, Pacchetti C, Blandini F. Amroxol-induced rescue of defective glucocerebrosidase is associated with increased LIMP-2 and saposin C levels in GBA1 mutant Parkinson's disease cells. *Neurobiol Dis*. 2015;82:235–42. doi:10.1016/j.nbd.2015.06.008.
 152. Siebert M, Sidransky E, Westbroek W. Glucocerebrosidase is shaking up the synucleinopathies. *Brain*. 2014;137:1304–22. doi:10.1093/brain/awu002.
 153. Weiser T. Amroxol: a CNS drug? *CNS Neurosci Ther*. 2008;14:17–24. doi:10.1111/j.1527-3458.2007.00032.x.
 154. Luan Z, Li L, Higaki K, Nanba E, Suzuki Y, Ohno K. The chaperone activity and toxicity of amroxol on Gaucher cells and normal mice. *Brain Dev*. 2013;35:317–22. doi:10.1016/j.braindev.2012.05.008.
 155. Khanna R, Benjamin ER, Pellegrino L, Schilling A, Rigat BA, Soska R, et al. The pharmacological chaperone isofagomine increases the activity of the Gaucher disease L444P mutant form of beta-glucosidase. *FEBS J*. 2010;277:1618–38. doi:10.1111/j.1742-4658.2010.07588.x.
 156. Steet RA, Chung S, Wustman B, Powe A, Do H, Kornfeld SA. The iminosugar isofagomine increases the activity of N370S mutant acid beta-glucosidase in Gaucher fibroblasts by several mechanisms. *Proc Natl Acad Sci U S A*. 2006;103:13813–8.
 157. Sun Y, Liou B, Xu YH, Quinn B, Zhang W, Hamler R, et al. Ex vivo and in vivo effects of isofagomine on acid beta-glucosidase variants and substrate levels in Gaucher disease. *J Biol Chem*. 2012;287:4275–87. doi:10.1074/jbc.M111.280016.
 158. Yang C, Rahimpour S, Lu J, Pacak K, Ikejiri B, Brady RO, et al. Histone deacetylase inhibitors increase glucocerebrosidase activity in Gaucher disease by modulation of molecular chaperones. *Proc Natl Acad Sci U S A*. 2013;110:966–71. doi:10.1073/pnas.
 159. Richter F, Fleming SM, Watson M, Lemesse V, Pellegrino L, Ranes B, et al. A GCase chaperone improves motor function in a mouse model of synucleinopathy. *Neurotherapeutics*. 2014;11:840–56. doi:10.1007/s13311-014-0294-x.
 160. Patnaik S, Zheng W, Choi JH, Motabar O, Southall N, Westbroek W, et al. Discovery, structure-activity relationship, and biological evaluation of noninhibitory small molecule chaperones of glucocerebrosidase. *J Med Chem*. 2012;55:5734–48. doi:10.1021/jm300063b.

161. Aflaki E, Stubblefield BK, Maniwang E, Lopez G, Moaven N, Goldin E, et al. Macrophage models of Gaucher disease for evaluating disease pathogenesis and candidate drugs. *Sci Transl Med*. 2014;6:240ra–73ra. doi:10.1126/scitranslmed.3008659.
162. Aflaki E, Borger DK, Moaven N, Stubblefield BK, Rogers SA, Patnaik S, et al. A New Glucocerebrosidase Chaperone Reduces alpha-Synuclein and Glycolipid Levels in iPSC-Derived Dopaminergic Neurons from Patients with Gaucher Disease and Parkinsonism. *J Neurosci*. 2016;36:7441–52. doi:10.1523/JNEUROSCI.0636-16.2016.
163. Xilouri M, Brekk OR, Landeck N, Pitychoutis PM, Papisilekas T, Papadopoulou-Daifoti Z, et al. Boosting chaperone-mediated autophagy in vivo mitigates alpha-synuclein-induced neurodegeneration. *Brain*. 2013;136:2130–46. doi:10.1093/brain/awt131.
164. Anguiano J, Garner TP, Mahalingam M, Das BC, Gavathiotis E, Cuervo AM. Chemical modulation of chaperone-mediated autophagy by retinoic acid derivatives. *Nat Chem Biol*. 2013;9:374–82. doi:10.1038/nchembio.1230.
165. East DA, Campanella M. Mitophagy and the therapeutic clearance of damaged mitochondria for neuroprotection. *Int J Biochem Cell Biol*. 2016;79:382–7. doi:10.1016/j.biocel.2016.08.019.
166. East DA, Fagiani F, Crosby J, Georgakopoulos ND, Bertrand H, Schaap M, et al. PMI: a DeltaPsim independent pharmacological regulator of mitophagy. *Chem Biol*. 2014;21:1585–96. doi:10.1016/j.chembiol.2014.09.019.
167. Jegga AG, Schneider L, Ouyang X, Zhang J. Systems biology of the autophagy-lysosomal pathway. *Autophagy*. 2011;7:477–89. doi:10.4161/auto.7.5.14811.
168. Kanagaraj N, Beiping H, Dheen ST, Tay SS. Downregulation of miR-124 in MPTP-treated mouse model of Parkinson's disease and MPP iodide-treated MN9D cells modulates the expression of the calpain/cdk5 pathway proteins. *Neuroscience*. 2014;272:167–79. doi:10.1016/j.neuroscience.2014.04.039.
169. Wang H, Ye Y, Zhu Z, Mo L, Lin C, Wang Q, et al. MiR-124 regulates apoptosis and autophagy process in MPTP model of Parkinson's disease by targeting to Bim. *Brain Pathol*. 2016;26:167–76. doi:10.1111/bpa.12267.
170. Alvarez-Erviti L, Seow Y, Schapira AH, Rodriguez-Oroz MC, Obeso JA, Cooper JM. Influence of micromRNA deregulation on chaperone-mediated autophagy and alpha-synuclein pathology in Parkinson's disease. *Cell Death Dis*. 2013;4:e545. doi:10.1038/cddis.2013.73.
171. Wu H, Chen S, Ammar AB, Xu J, Wu Q, Pan K, et al. Crosstalk Between Macroautophagy and Chaperone-Mediated Autophagy: Implications for the Treatment of Neurological Diseases. *Mol Neurobiol*. 2014;52(3):1284–96. doi:10.1007/s12035-014-8933-0.
172. Tanik SA, Schultheiss CE, Volpicelli-Daley LA, Brunden KR, Lee VM. Lewy body-like alpha-synuclein aggregates resist degradation and impair macroautophagy. *J Biol Chem*. 2013;288:15194–210. doi:10.1074/jbc.M113.457408.

Submit your next manuscript to BioMed Central and we will help you at every step:

- We accept pre-submission inquiries
- Our selector tool helps you to find the most relevant journal
- We provide round the clock customer support
- Convenient online submission
- Thorough peer review
- Inclusion in PubMed and all major indexing services
- Maximum visibility for your research

Submit your manuscript at
www.biomedcentral.com/submit

